

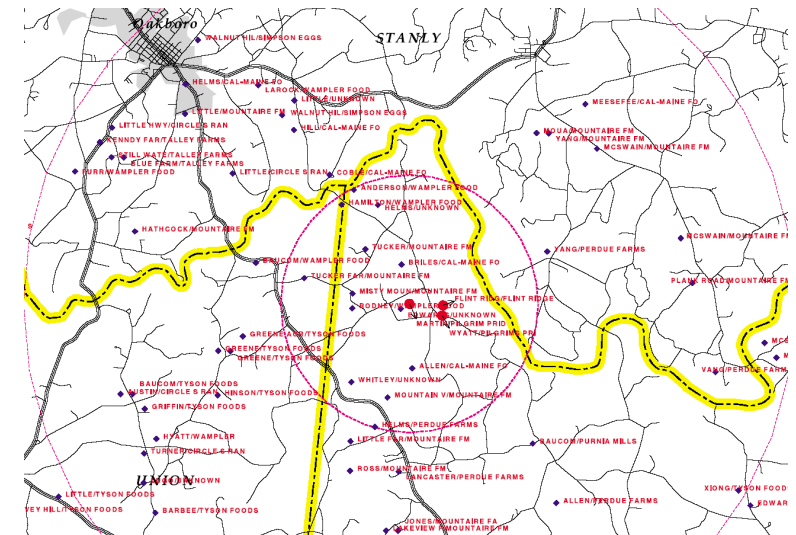
The neglected components of biosecurity programs



What works
Why it often does not
What we can do about it

Jean-Pierre Vaillancourt

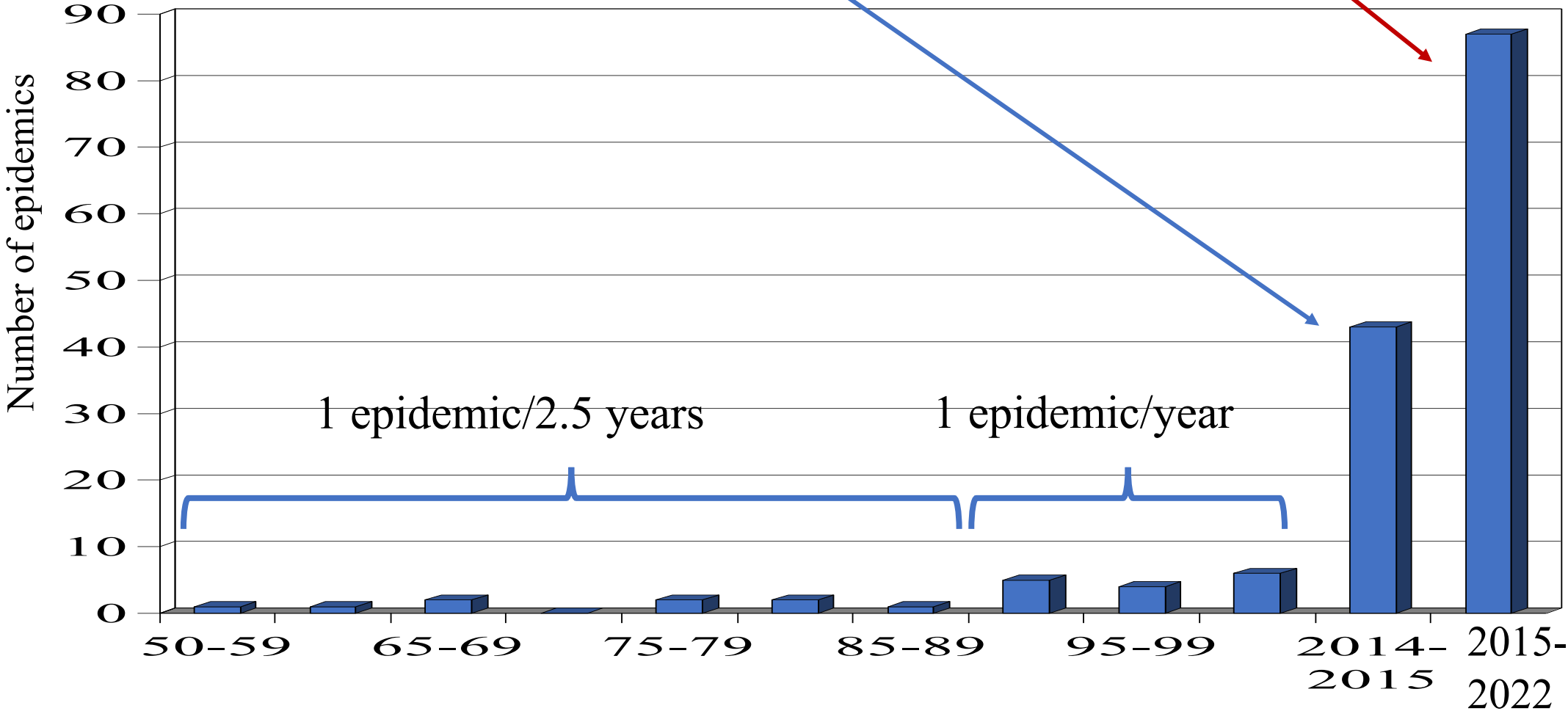
Université 
de Montréal



Highly Pathogenic Avian Influenza

43 H5 & H7 epidemics; 7 different viruses in 22 countries in Africa, Americas, Asia, Australia, Europe, Middle East

Over 80 H5 epidemics; over 10 strains in over 75 countries



Avian Influenza Viruses With Zoonotic Potential

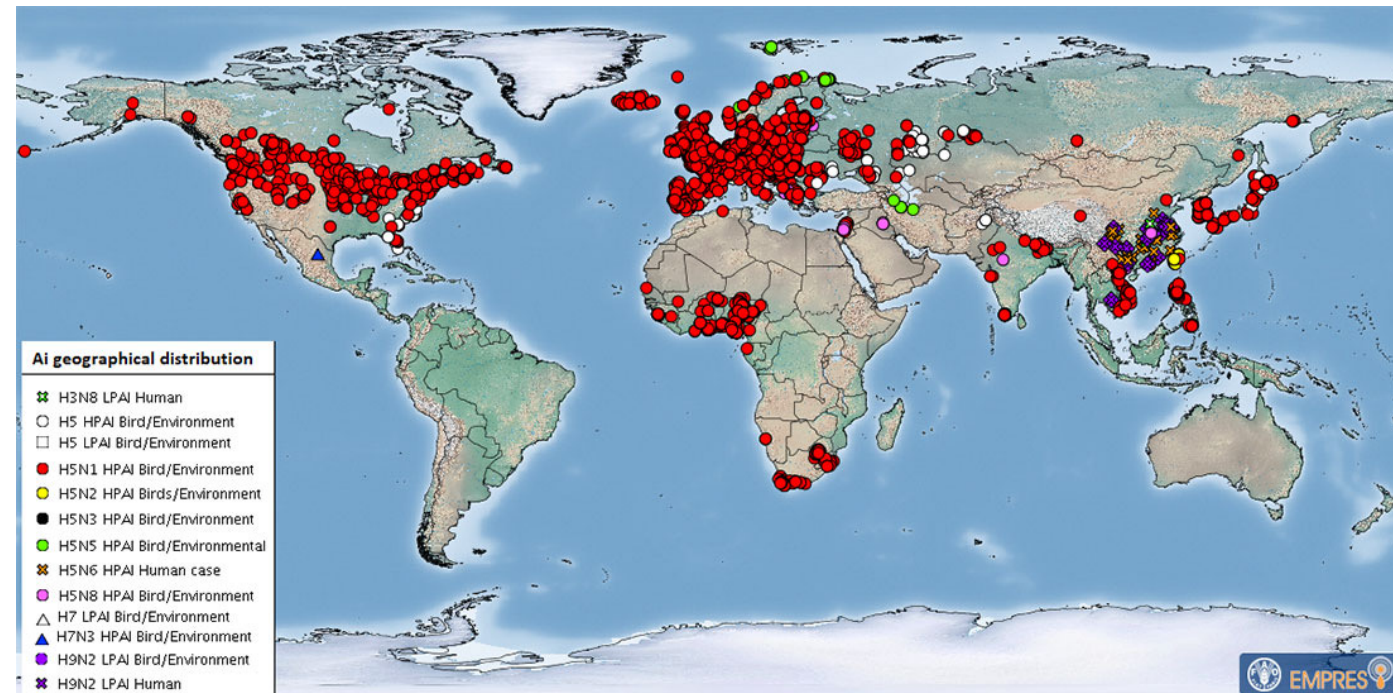
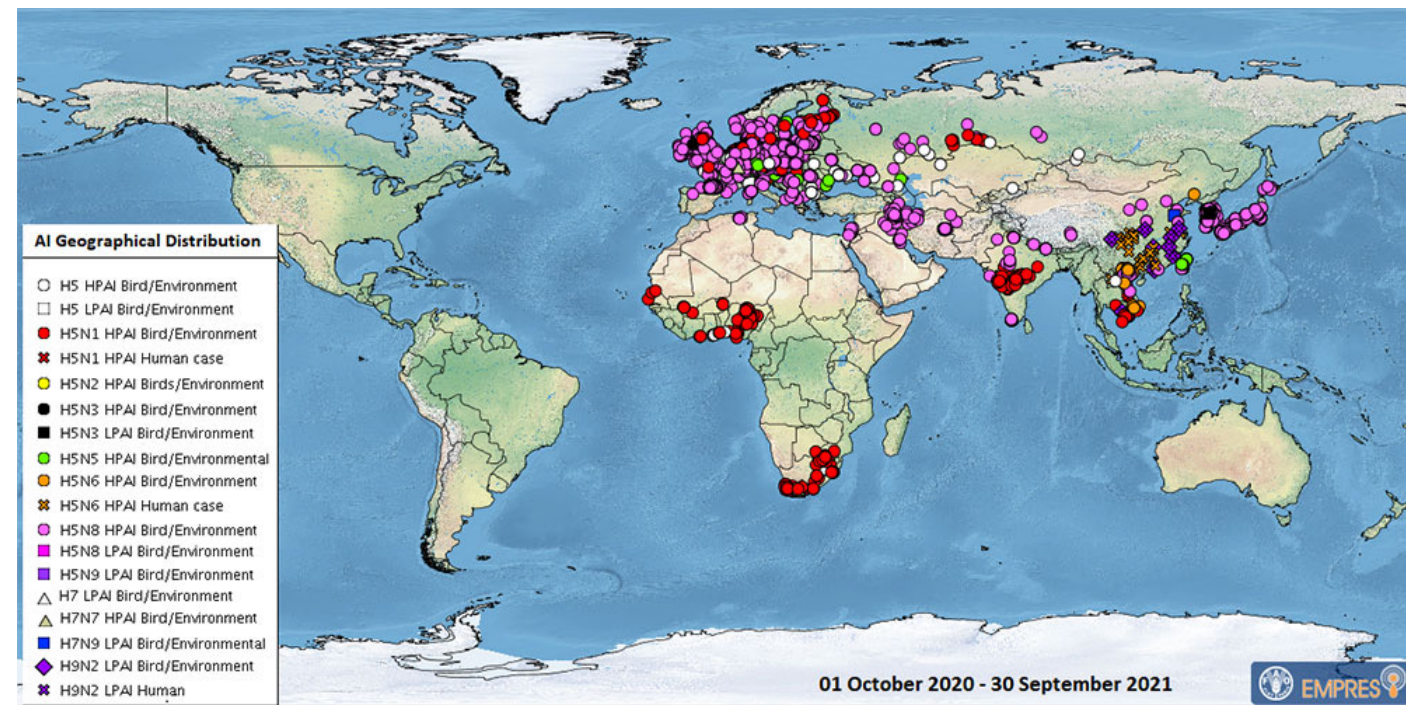
October 2020 to
September 2021 →

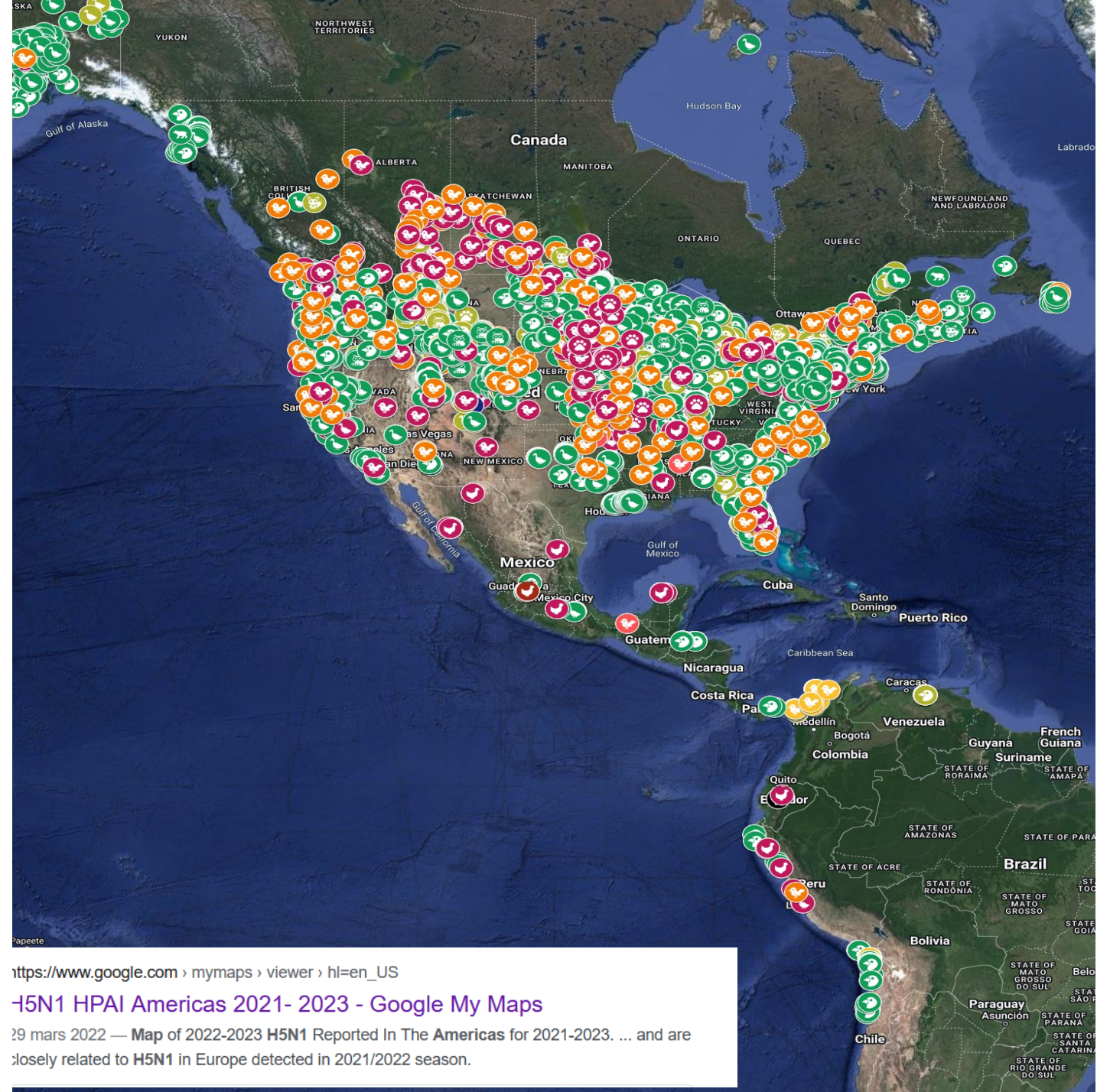
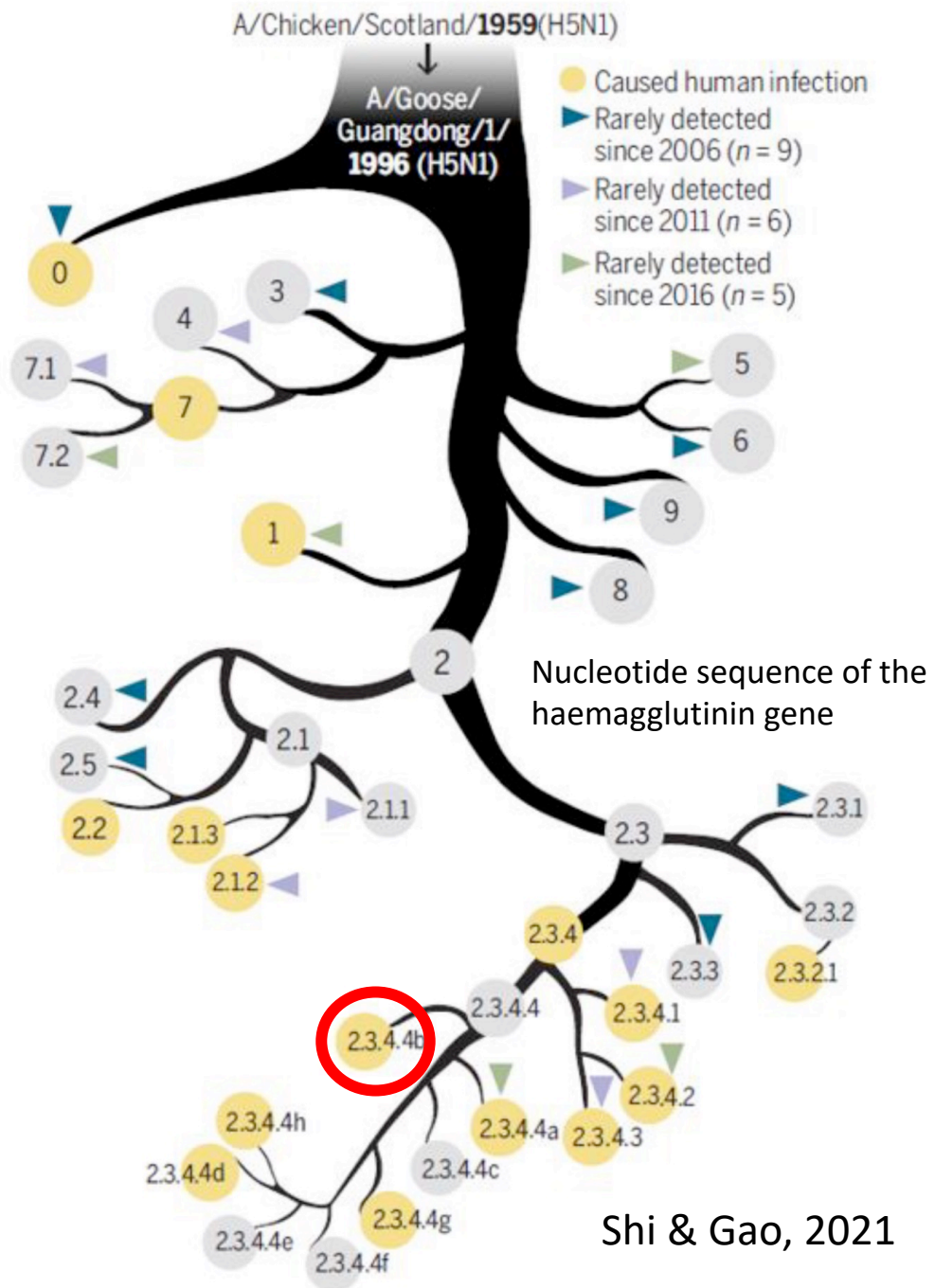
H5N8

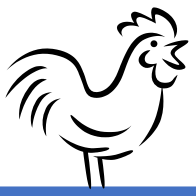
Dominance

October 2021 to
September 2022 →

H5N1







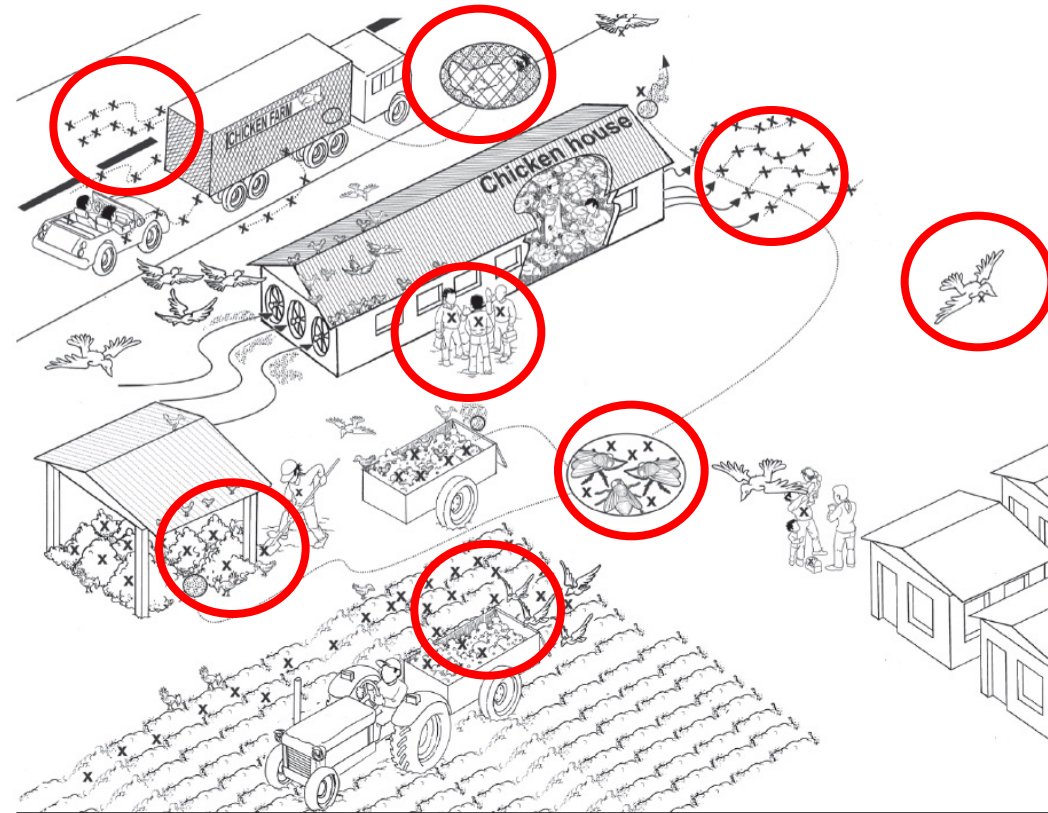
Risk factors associated with Avian Influenza

Country	Species	Significant Factors	Reference
USA	Turkey Chicken	Odds are 7.3 times higher when disposing dead birds by <u>rendering</u>	McQuiston et al., 2005
Japan	Layer	Odds are 37 times higher when introducing end-of-lay chickens Odds are 29.4 times higher when <u>sharing farm equipment</u> between farms Odds are 7 times higher when visitors have incomplete hygiene measures (<u>shoes, clothes hands</u>)	Nishiguchi et al., 2007
Korea	Layer	Odds are 500 times lower when farms have disinfection stations (hand & boot washing/disinfection) Odds are 6 times <u>lower when changing boots between barns</u>	Yoo et al. 2002
France	Duck	Odds are 9 times higher when inadequate management of vehicles and people movements Odds are 6.5 times higher if inadequate delimitation of farm and units (leading to inadequate use of anteroom)	Guinat et al., 2020
Netherlands	All	Odds are 2 times higher in layer compared to other types of birds	Thomas et al., 2005
Italy	All	Farms located within 1500 m from case farms are 12 times more likely to be infected. Turkeys are 4 times more likely to be infected compared to other species.	Mannelli et al., 2006

Risk identification – we know the risks

- Birds
- Multi-species sites
- Water
- Visitors & employees
- Equipment
- Dead bird disposal
- Manure management
 - Storage
 - Spread
- Rodents & other wildlife
- Insects
- Regional farm density
- The wind?

Figure 4. A schematic representation (not to scale) of multiple potential pathways for exposure to and transfer of pathogens within the environs of concentrated animal feeding operations*



(Artist: Salvador Saenz)

*Compromises to biosecurity include: (1) workers lacking protective clothing or opportunities for personal hygiene or decontamination on-site; (2) inadequate management of animal biosolids, often applied to land without treatment; (3) flies and other insects that carry pathogens in and out of facilities through ventilation systems and small openings; (4) ventilation with high-volume fans resulting in considerable movement of materials into the external environment; and (5) transporting animals in open trucks or containers to the farm or for processing.

DEUTERONOMY



“You shall have a place outside the camp, and you shall go out to it. And you shall have a trowel with your tools, and when you sit down outside, you shall dig a hole with it and turn back and cover up your excrement.”

“And he that is to be cleansed shall wash his clothes, ...wash himself in water, ...and after that he shall come into the camp, but shall stay outside of his tent seven days.”

Leviticus/14/8

Soldiers returning from war were required to flame all their equipment and to plunge garments in boiling water.



DEUTERONOMY



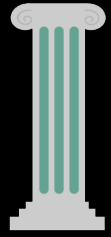
“You shall have a place outside the camp, and you shall go out to it. And you shall have a trowel with your tools, and when you sit down outside, you shall dig a hole with it and turn back and cover up your excrement.”

- ✓ “I forbid you to ever enter a ...an assembly of people.
- ✓ I forbid you to leave your house unless dressed in your recognizable garb
- ✓ I forbid you to drink at any stream or fountain, unless using your own barrel
- ✓ I forbid you to touch anything...until it becomes your own.
- ✓ I forbid you to share house with any woman but your wife.
- ✓ If accosted by anyone...set yourself downwind of them
- ✓ I forbid you to enter any narrow passage, lest a passerby bump into you.
- ✓ I forbid you...to touch anything without donning your gloves.
- ✓ I forbid you to drink or eat from any vessel but your own.”

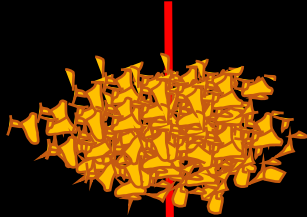
**Separatio Leprosorum
(Mass of separation)**



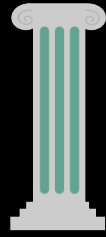
Basic Biosecurity Principles



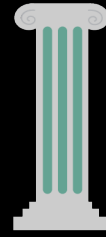
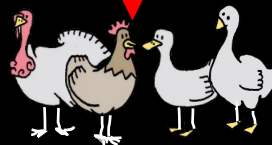
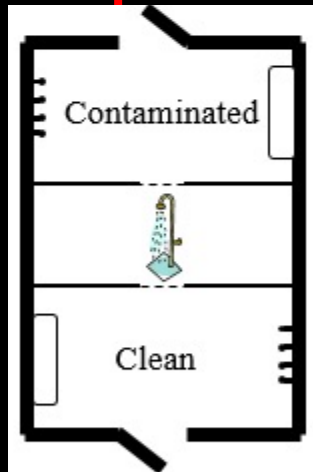
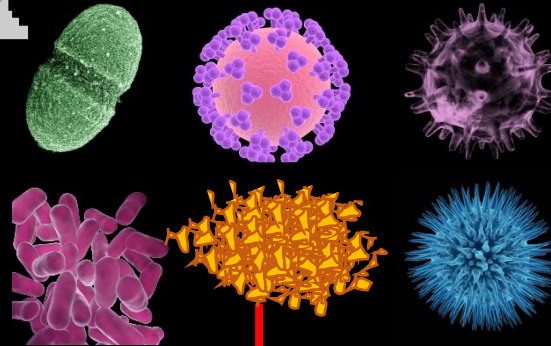
Reduce



Clean



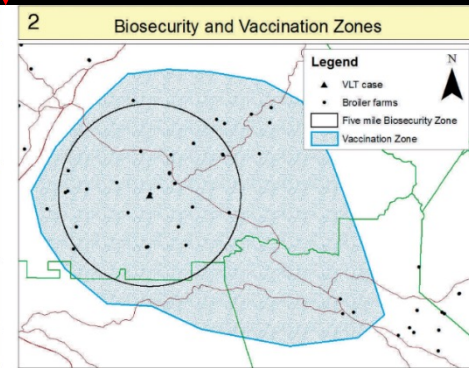
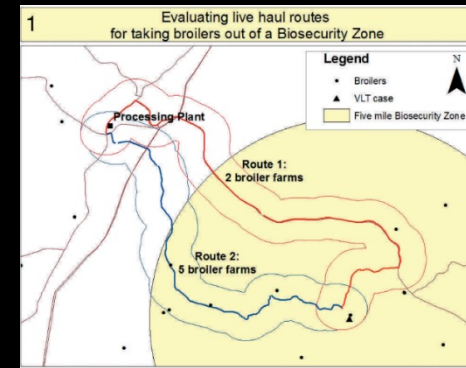
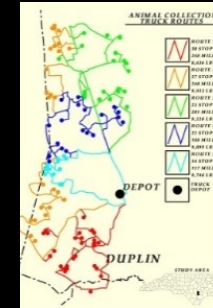
Separate



Communicate



Organize



Cleaning & Disinfection

- Inadequate contact time (detergent and disinfectant)
- Failure to use soap/detergent when cleaning
- Failure to get everything dry before disinfecting
- Disinfecting immediately after cleaning rather than just prior to use (equipment being stored outside after C&D and getting contaminated with bird droppings)
- Monitoring C&D (changes in personnel, interpretation differences on “clean”, etc.)
- Failure to observe the process
- Failure to make sure C&D and biosecurity supplies are being ordered and available



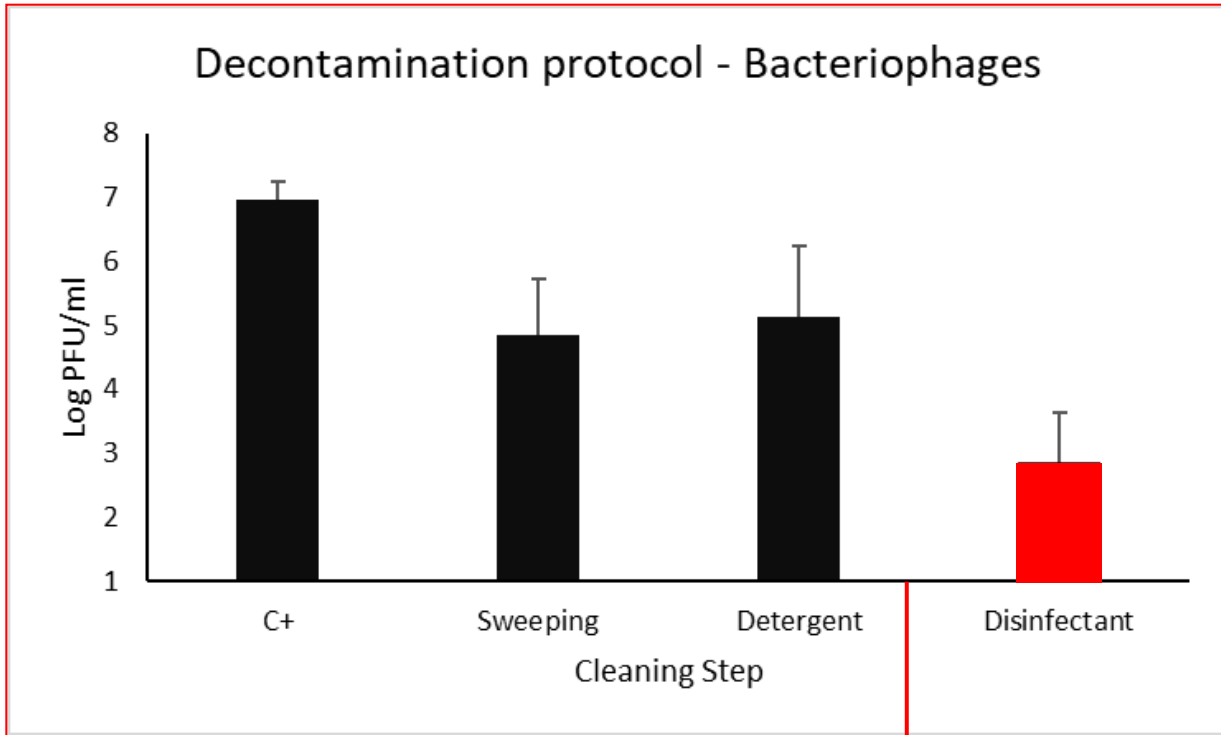
Gonder, 2023



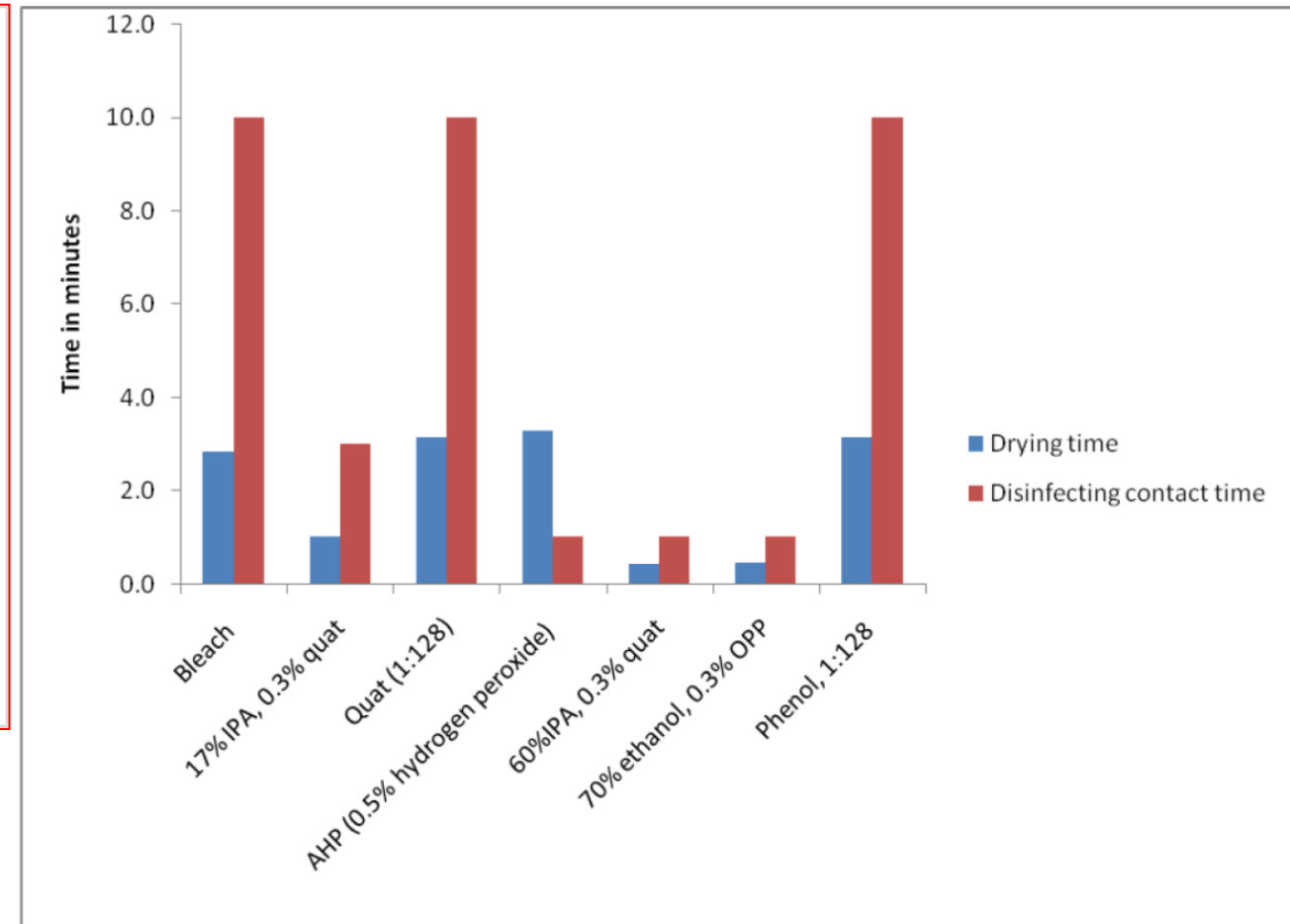
Impact of drying and disinfection on viruses



Table 1: Drying Time versus Disinfectant Label Contact Time



Drying



The Importance of Contact Times for Disinfectants
(Omidbakhsh, N. CJIC 2008; 23:49)

An assessment of sanitation protocols for commercial transport vehicles contaminated with porcine reproductive and respiratory syndrome virus

Scott Dee, John Deen, Danny Burns, George Douthit, Carlos Pijoan

Table II. Summary of diagnostic data from

Area tested	Wash only	and	Wash + Formaldehyde	Trt 4	Neg Ctrl	
Trailer interior pretreatment	20/20 ^a		20/20	19/20	20/20	0/20
Trailer interior 60 min posttreatment	20/20		20/20	2/19	NT	0/20
Trailer interior 90 min posttreatment	20/20		20/20	0/19	NT	0/20
Trailer interior allowed to dry (8 h)	NT		NT	NT	0/20	0/20
Number of PRRSV (+) pigs postexposure	2/4 ^b		2/4	0/4	0/4	0/4

Trt 1 — Treatment 1, washing only; Trt 2 — washing plus glutaraldehyde: quaternary ammonium chloride; Trt 3 — washing plus glutaraldehyde: quaternary ammonium chloride and overnight drying; Neg Ctrl — Sham-inoculated protocol

Wash +
Glutaraldehyde: quaternary ammonium chloride

Trt 3 — Treatment 3, washing plus glutaraldehyde: quaternary ammonium chloride and overnight drying

^a Number of polymerase chain reaction (PCR)-positive swabs per number of replicates conducted

^b Number of replicates that demonstrated PRRSV infection of naïve sentinel pigs housed in the trailer for 2 h per number of replicates conducted

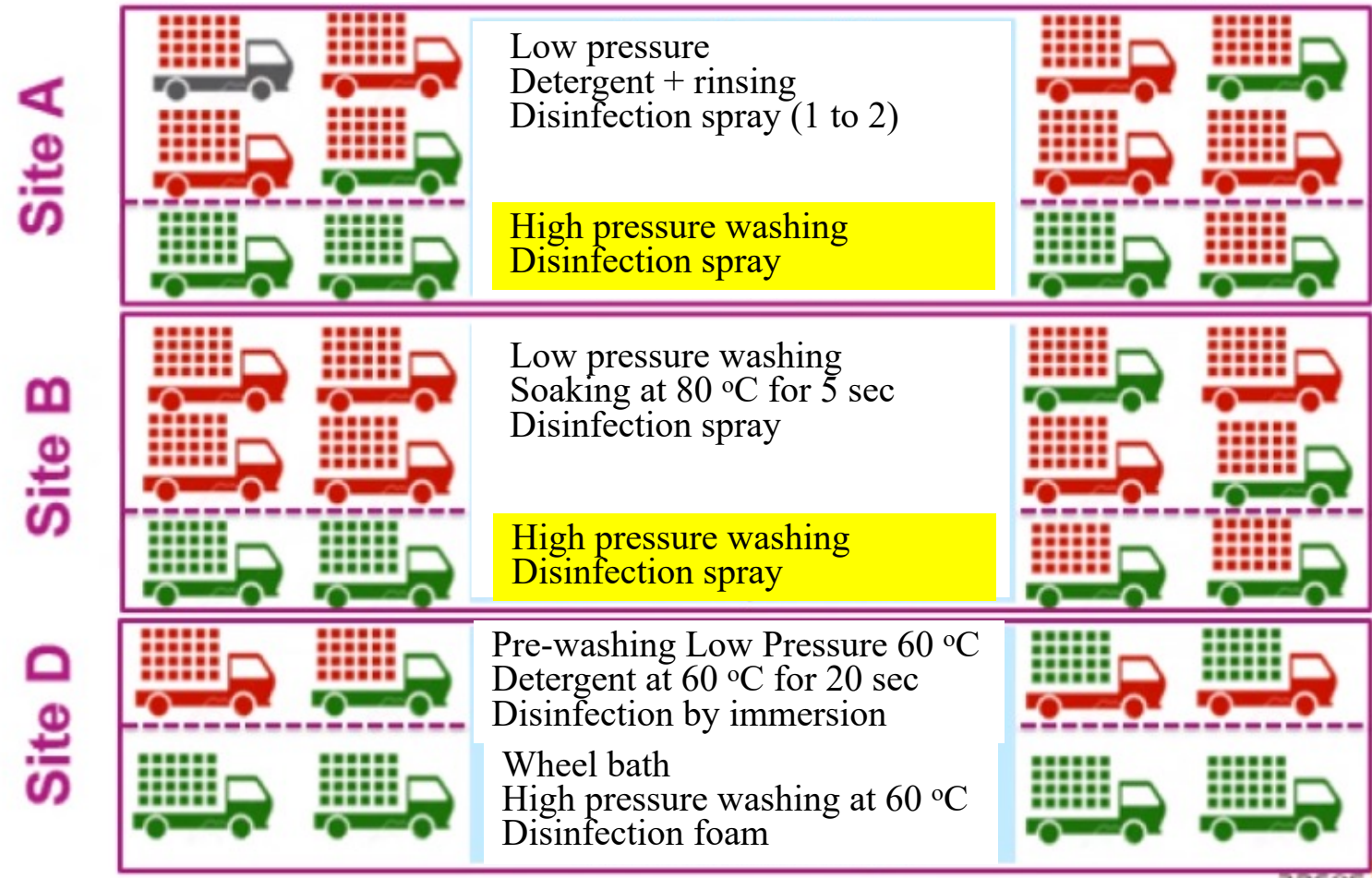
Wash +
Overnight drying

Cleaning and disinfection of crates and trucks used for duck transport: field observations during the H5N8 avian influenza outbreaks in France in 2017

Adeline Huneau-Salaün, Axelle Scoizec, Rodolphe Thomas, Sophie Le Bouquin

Before

After



■ Gene M+

■ Gene M-



Failure

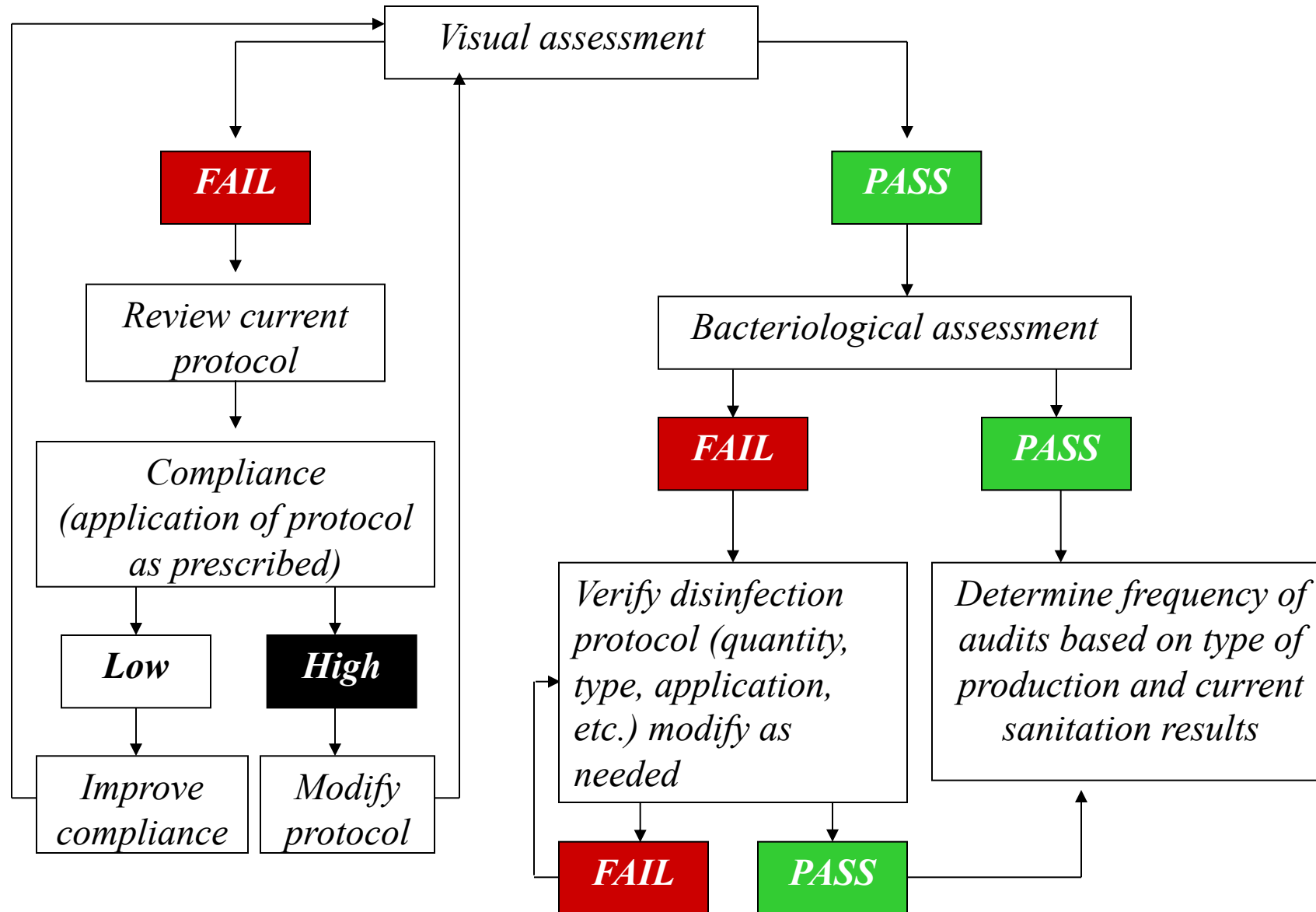
Initial amount of contamination

Poor execution

Cross-contamination after cleaning



Monitoring of sanitation

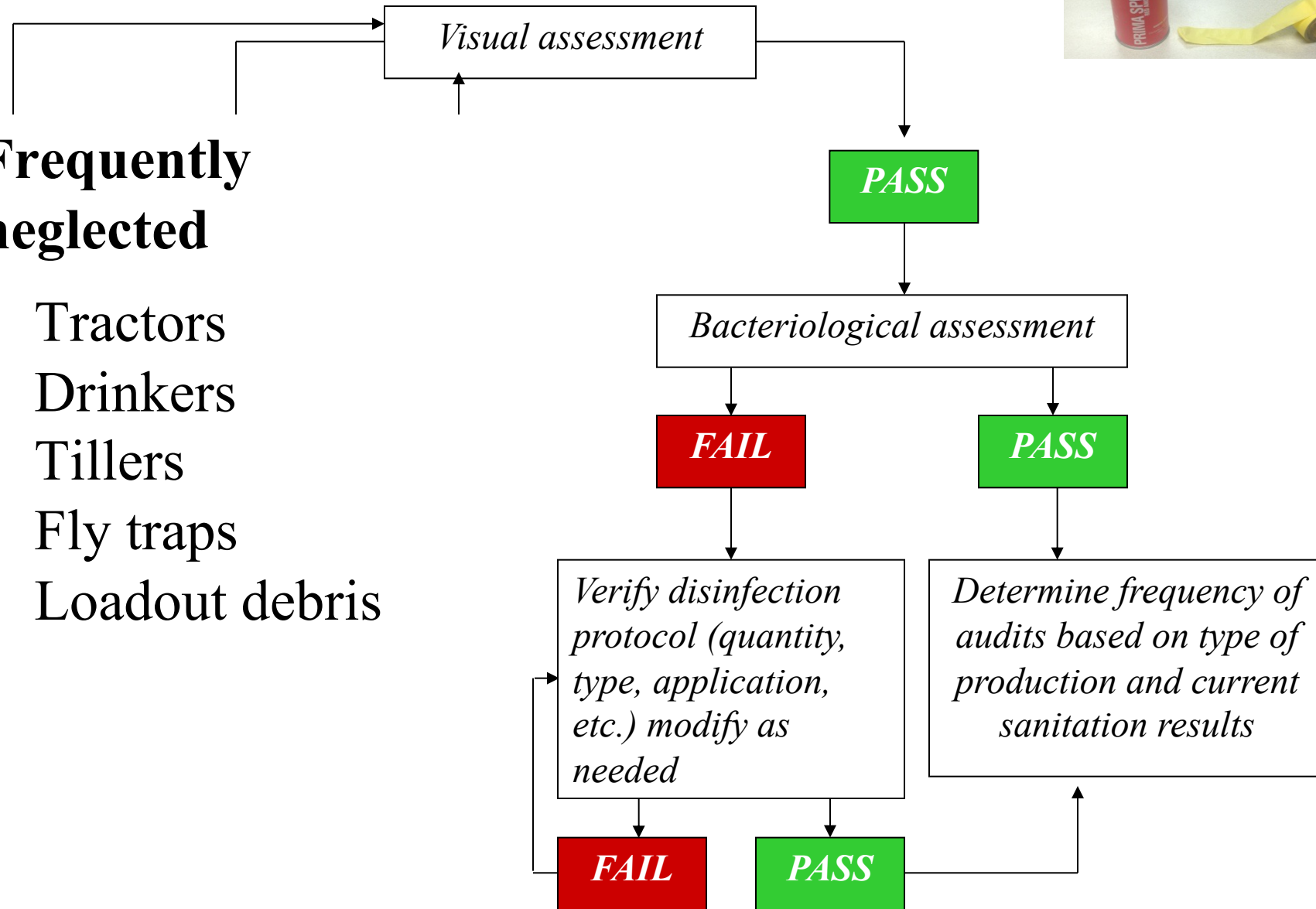


Mark the spots needing attention, especially if person responsible is not present



Frequently neglected

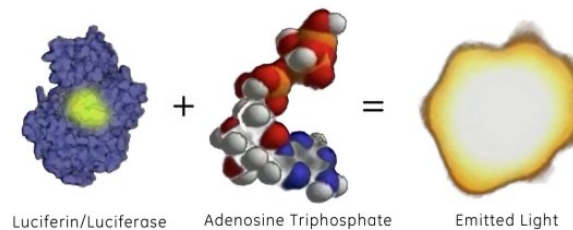
- Tractors
- Drinkers
- Tillers
- Fly traps
- Loadout debris



Colorimetric representation of important attributes for a successful microbial monitoring system

Detection method	Accurate	Rapid	Cost-effective	Commercially practical
Agar air plate	Yellow	Yellow	Green	Green
RODAC plate	Green	Yellow	Green	Green
Direct swab	Green	Yellow	Green	Green
Fluff evaluation	Green	Red	Yellow	Yellow
MPN enumeration	Green	Red	Red	Yellow
ATP bioluminescence	Green	Green	Green	Blue

-  Positive
-  Neutral
-  Negative
-  Unknown



<https://www.hygiena.com/>



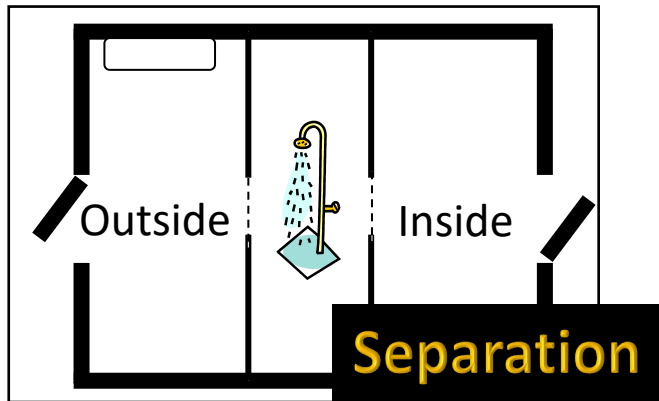
France



Mexico



Ethiopia 3/2018



USA



Cameroon 08/23/20



France

Study on the usage of footbaths under field conditions



Farms

Dry footbath

Total Bacterial Counts from Shoe Swabs

	Fresh Solution	After 3 hours of use
Active Ingredient	% Change in bacterial count	% Change in bacterial count
Phenol	-45.8	+130.5
Quaternary Ammonium	-57.5	+73.3
Water	+87.2	+44.8

Active Ingredient	% Change in Bacterial count	Average Residual Life
Detergent + Dry Bleach	-92.6	14 days
Dry Bleach	-98.06	14 days
Phenol	+ 10.8	<2 hours
Quaternary ammonium	-23.6	<2 hours

Robert L. Owen and John L.

25. QUINN
In Disin:
Philadel



Study on the usage of footbaths under field conditions



Farms

Dry footbath

Total Bac
from Sho

Active Ingredient		
Phenol	-4	
Quaternary Ammonium	-5	
Water	+87.2	+44.8



25. QUINN
In Disin
Philadel

Robert L. Owen and John La

Persistence of Highly Pathogenic and Low Pathogenic Avian Influenza Viruses in Footbaths and Poultry Manure

R. Hauck,^A B. Crossley,^B D. Rejmanek,^B H. Zhou,^C and R. A. Gallardo^{AD}

Table 1. Detection of HPAI and LPAI by RT-qPCR and virus isolation in spiked bedding material scraped from boots treated with quaternary ammonia + glutaraldehyde–, quaternary ammonia only–, or bleach powder–based footbaths.

	Hours after preparation of footbath							
	0		24		48		72	
	HPAI	LPAI	HPAI	LPAI	HPAI	LPAI	HPAI	LPAI
Control (feces no disinfectant)								
RT-qPCR	+ ^A	+	+	+	+	+	+	+
Isolation	+	+	+	+	+	+	+	+
Quaternary ammonia + glutaraldehyde								
RT-qPCR	+	+	+	+	+	+	+	+
Isolation	+	+	+	+	+	+	+	+
Quaternary ammonia								
RT-qPCR	+	+	+	+	+	+	+	+
Isolation	+	+	+	+	+	+	+	+
Bleach powder								
RT-qPCR	– ^B	–	–	–	–	–	–	–
Isolation	–	–	–	–	–	–	–	–

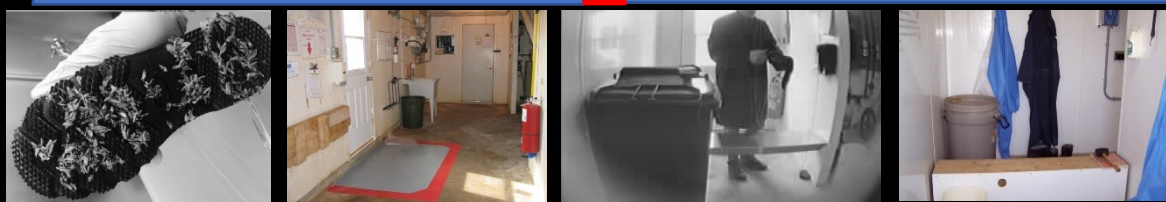
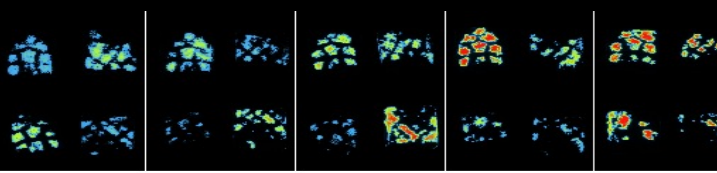
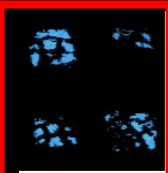
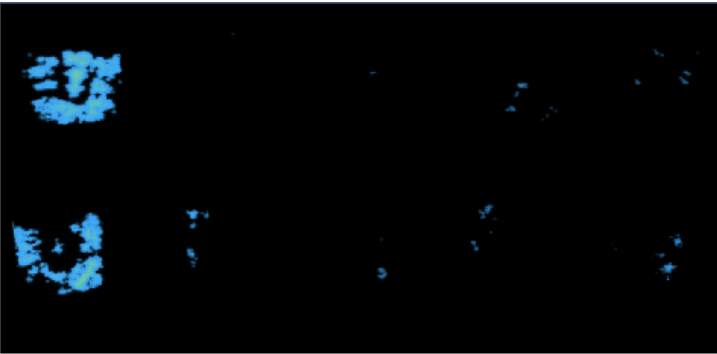
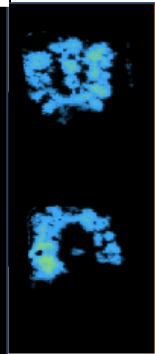
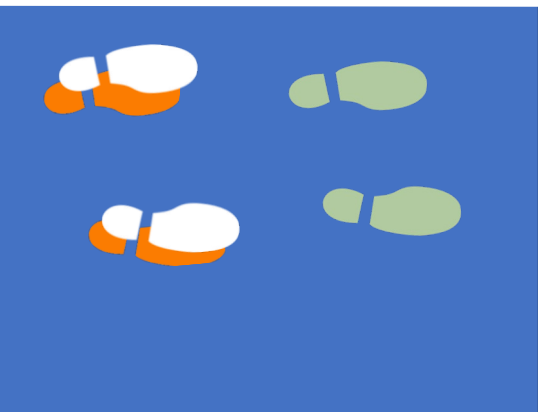
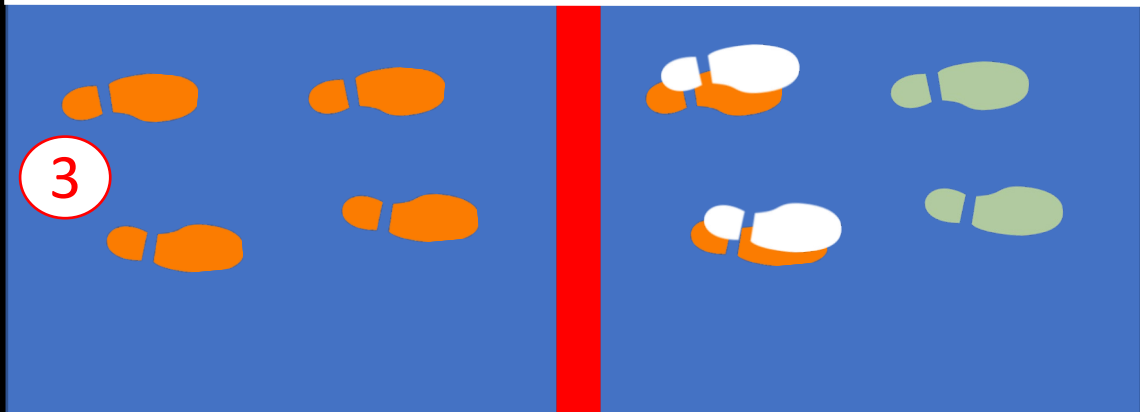
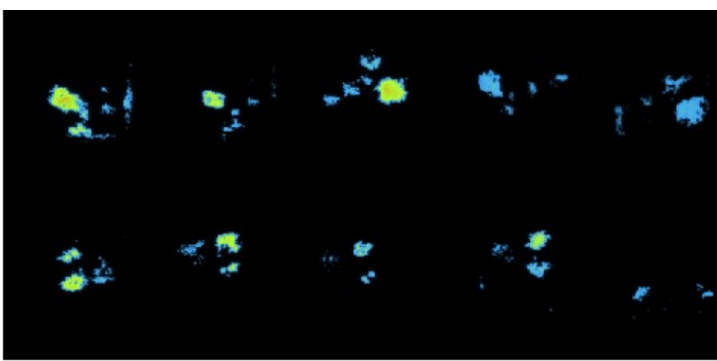
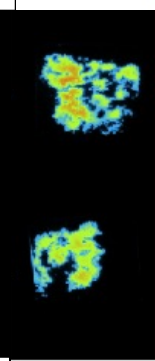
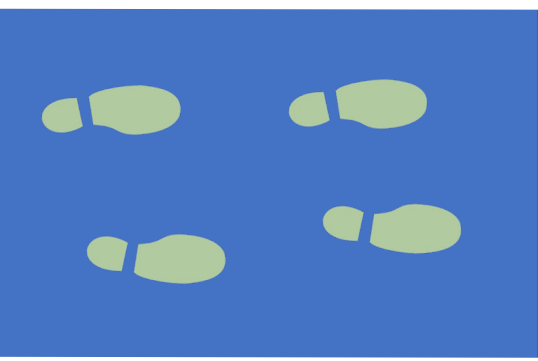
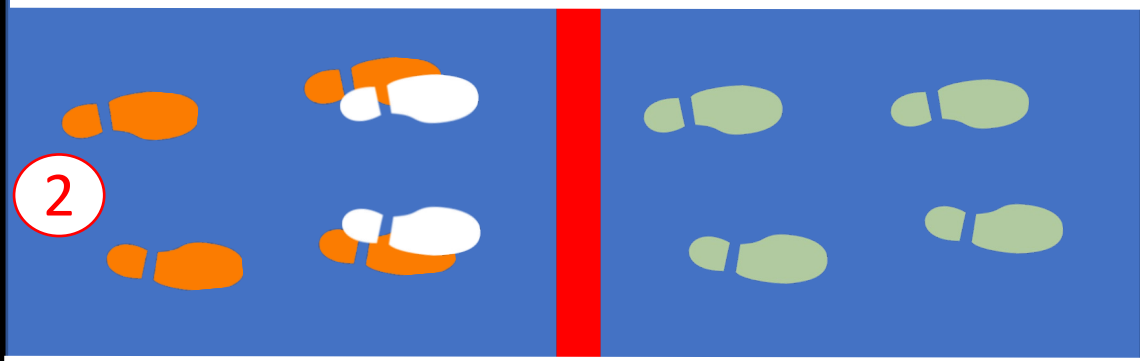
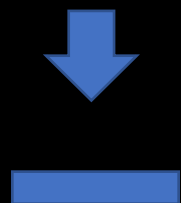
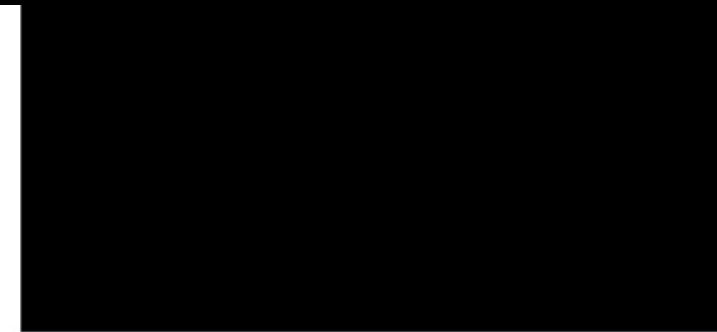
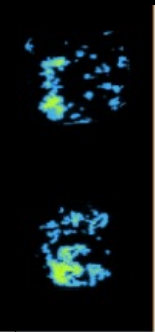
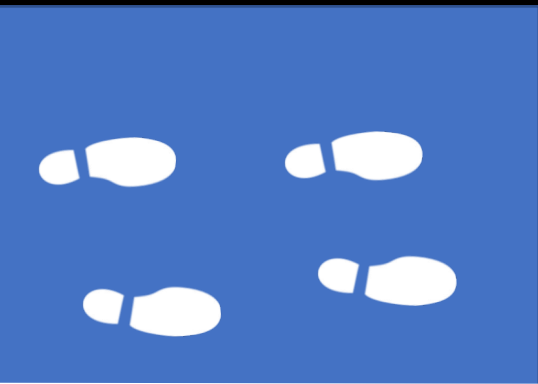
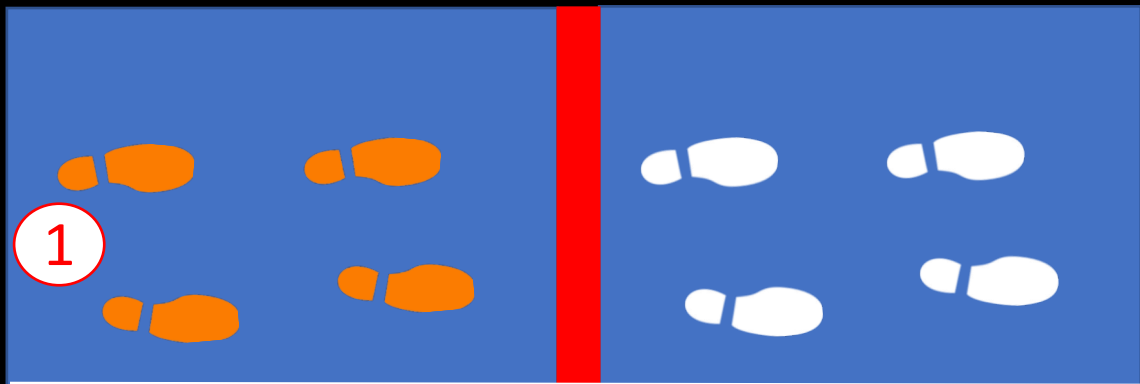
^AVirus detected.

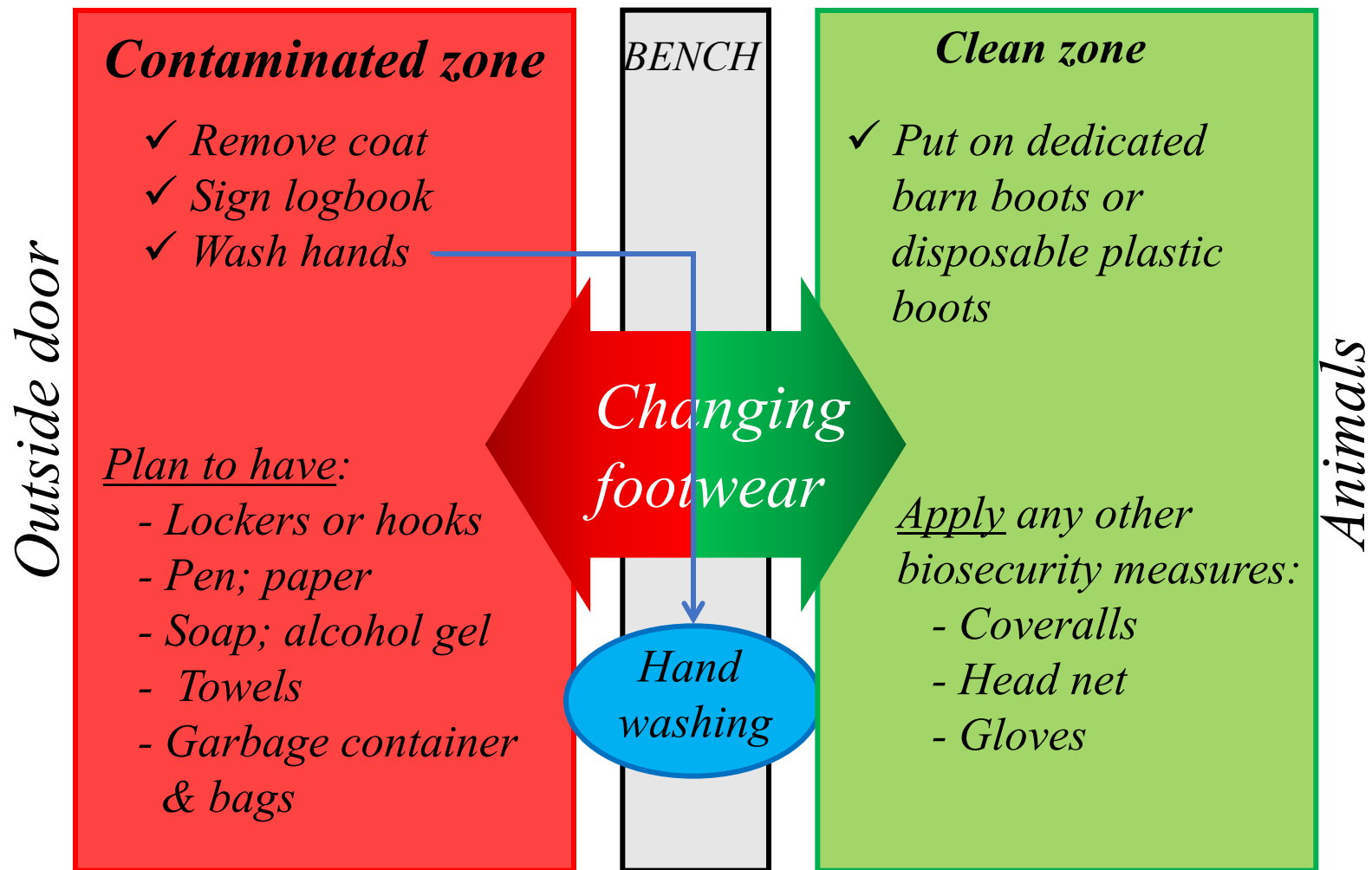
^BVirus not detected.



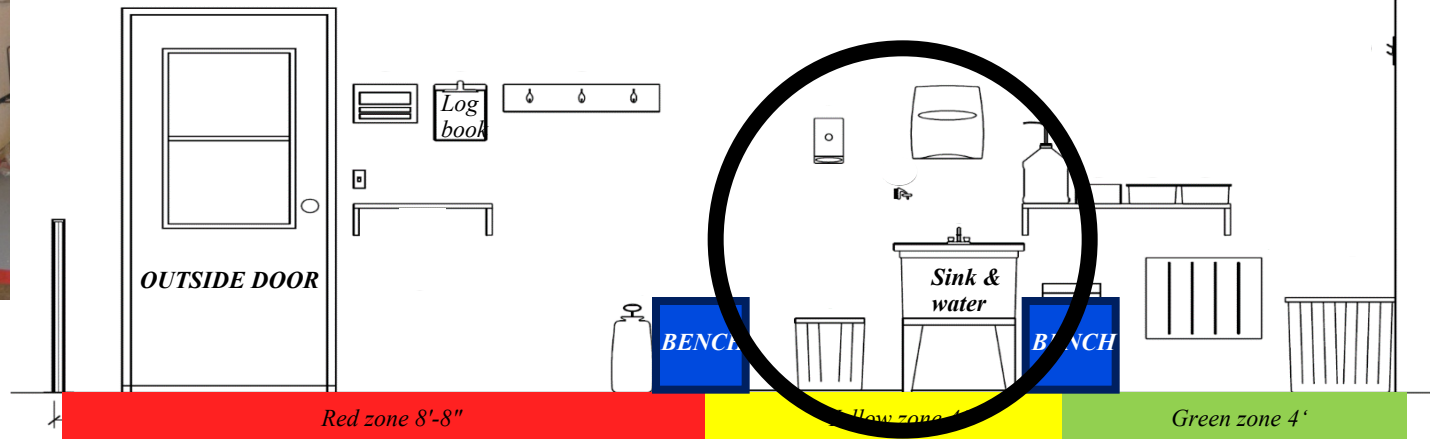
Fig. 1. (a) Manure accumulated in the boot crevices. (b) Sampling involved elimination of the excess of material in the surface and collection of the material inside the crevices.

Contamination





3-zone entrance



Outsi

Plan to have:

- Lockers or hooks
- Pen; paper
- Soap; alcohol gel
- Towels
- Garbage container & bags

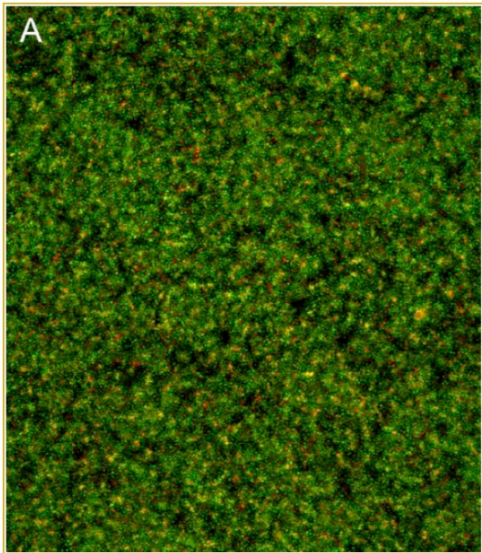
footwear



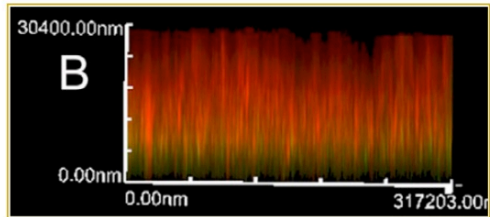
Apply any other biosecurity measures:

- Coveralls
- Head net
- Gloves

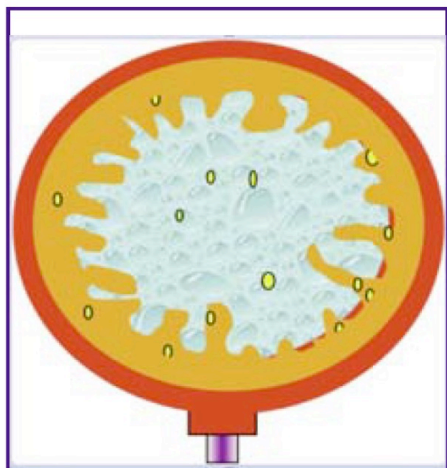
Anit.



Source : Y.D.N. Tremblay



Source : Y.D.N. Tremblay



SE Watkins



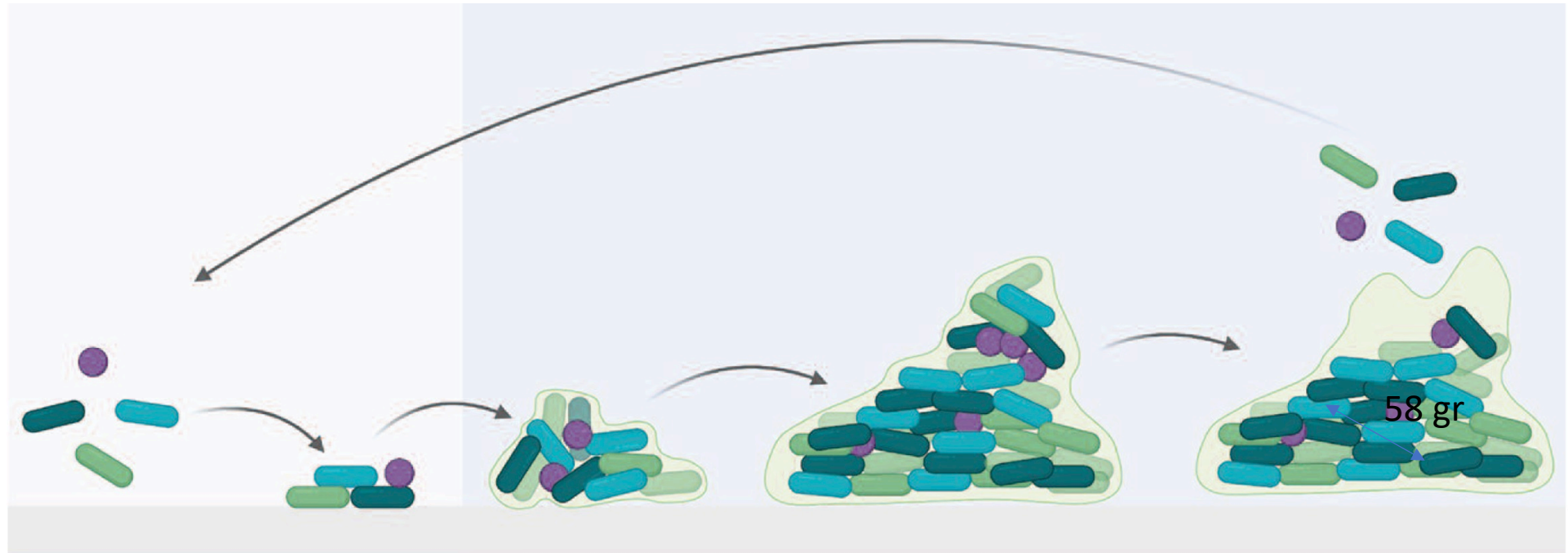
D Venne



D Venne



SE Watkins



Mobility

Getting to the surface

Adhesion

Adhesion to the surface

Growth

Matrix formation

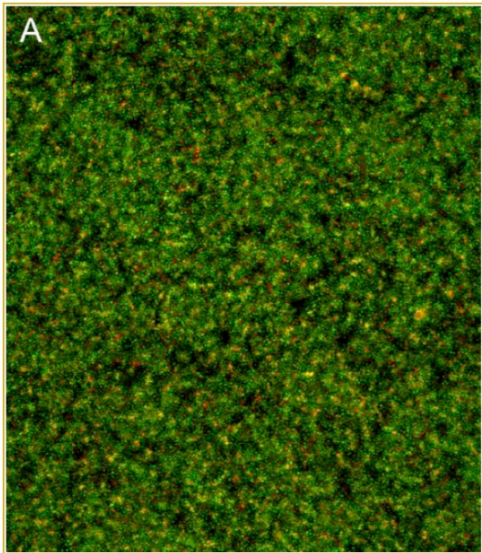
Maturation

Bacterial growth & development of the matrix

Dispersion

Start of a new cycle

58 gr



Source : Y.D.N. Tremblay



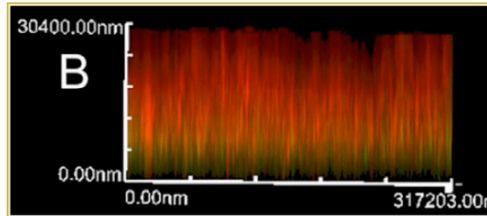
D Venne



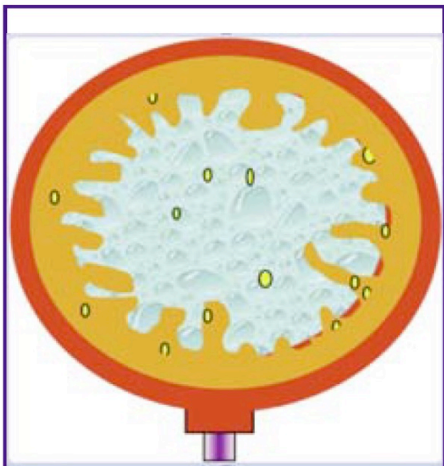
D Venne



SE Watkins



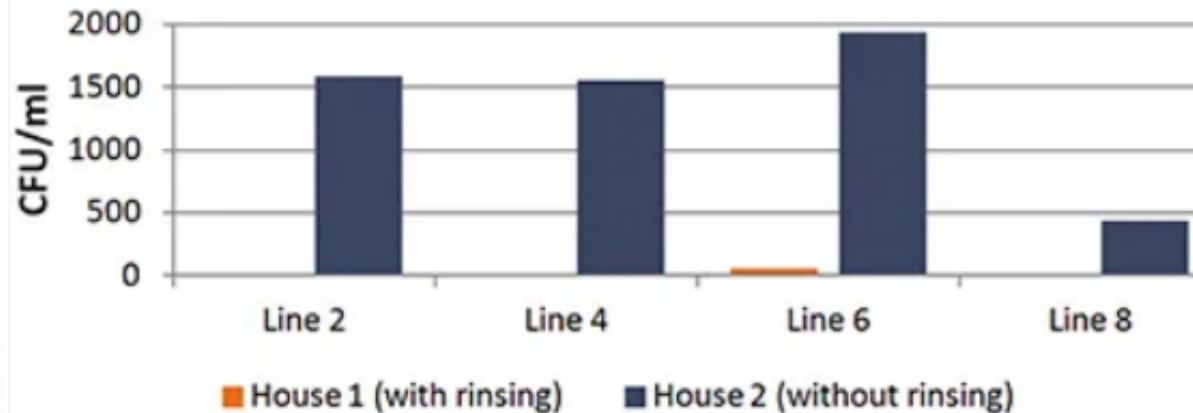
Source : Y.D.N. Tremblay



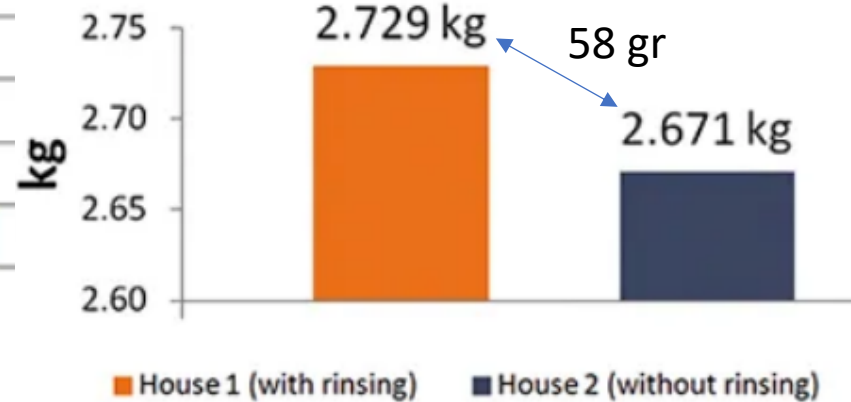
SE Watkins



Bacterial concentration: colony-forming units (CFU) at 22°C



Individual bird weights at moving-out



Biofilm formation in bacterial pathogens of veterinary importance

Mario Jacques^{1*}, Virginia Aragon^{2,3} and Yannick D. N. Tremblay¹

Bordetella

Campylobacter coli

Campylobacter jejuni

Clostridium perfringens

Enterococcus faecalis

Erysipelothrix rhusiopathiae

Escherichia coli

Listeria monocytogenes

Mycoplasmas

Pasteurella multocida

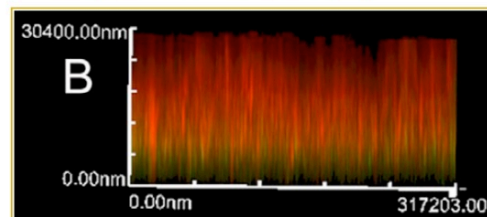
Pseudomonas aeruginosa

Riemerella anatipestifer

Samonella

Staphylococcus

Streptococcus

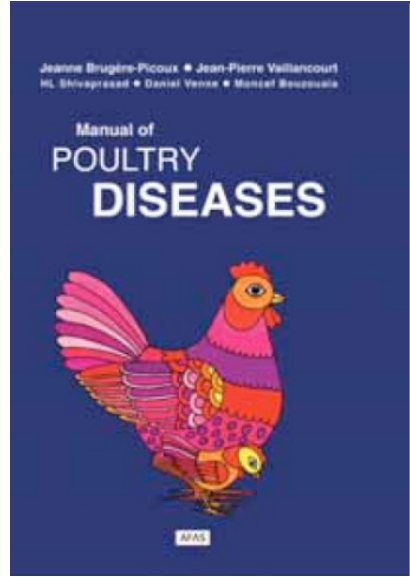
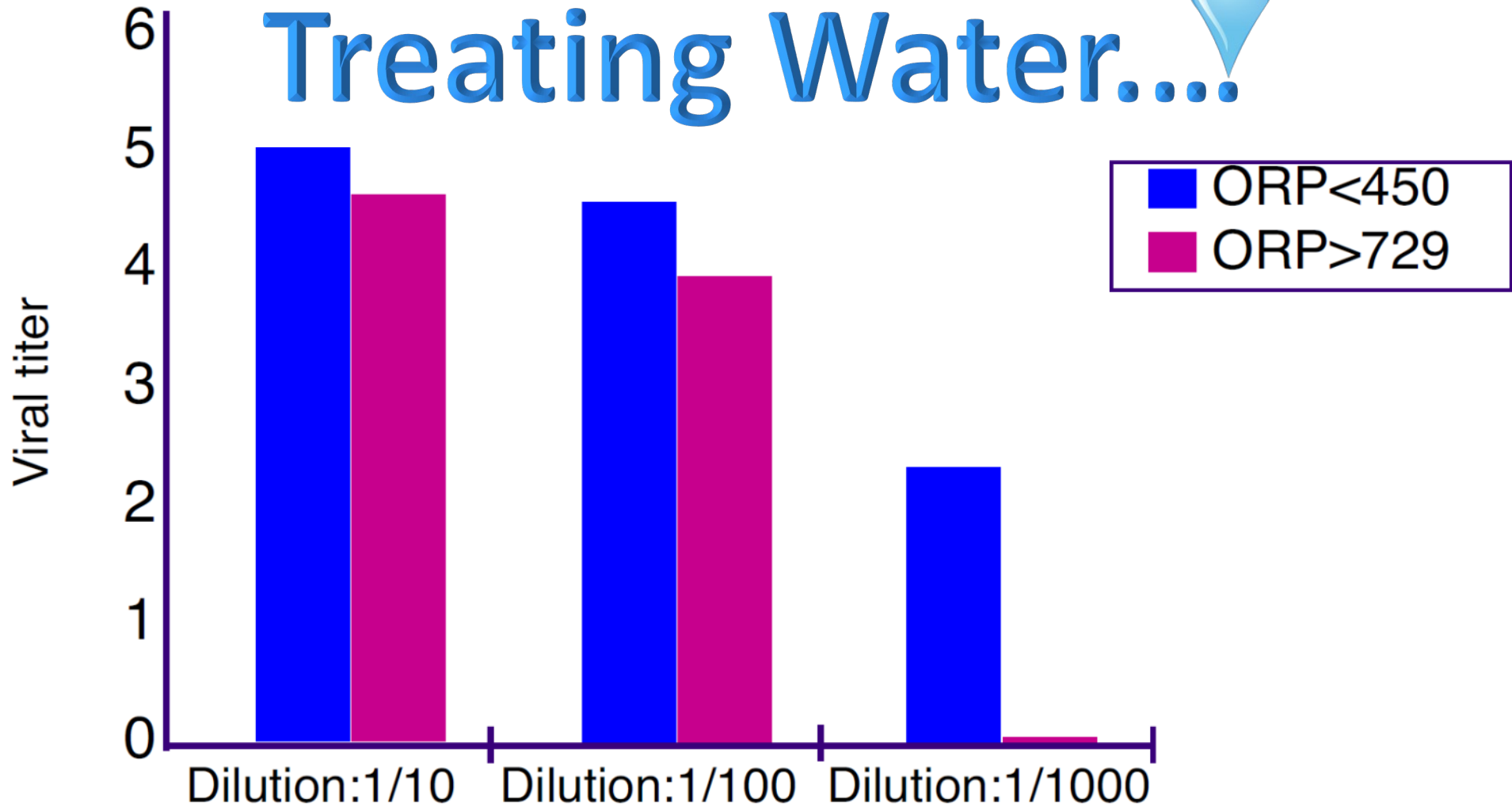


Source : Y.D.N. Tremblay

Table 1. Studies on biofilm formation of bacterial pathogens of veterinary importance

Bacterial species	References
<i>Actinobacillus pleuropneumoniae</i>	Auger <i>et al.</i> (2009a), Bossé <i>et al.</i> (2010), Buettner <i>et al.</i> (2008), Dalai <i>et al.</i> (2009), Ganeshnaryan <i>et al.</i> (2009), Izano <i>et al.</i> (2007), Kaplan <i>et al.</i> (2004), Kaplan and Mulks (2005), Kerrigan <i>et al.</i> (2008), Labrie <i>et al.</i> (2010), Li <i>et al.</i> (2008), Liu <i>et al.</i> (2008), Tegetmeyer <i>et al.</i> (2009)
<i>Aeromonas hydrophila</i>	Asha <i>et al.</i> (2004), Gavin <i>et al.</i> (2002), Kozlova <i>et al.</i> (2008), Lynch <i>et al.</i> (2002), Truchado <i>et al.</i> (2009)
<i>Arcanobacterium pyogenes</i>	Jost and Billington (2005), Olson <i>et al.</i> (2002)
<i>Bacillus cereus</i> group	Auger <i>et al.</i> (2006), Auger <i>et al.</i> (2009b), Houry <i>et al.</i> (2010), Lee <i>et al.</i> (2007), Schuch and Fischetti (2009), Shaheen <i>et al.</i> (2010), Shi <i>et al.</i> (2004), Wijman <i>et al.</i> (2007)
<i>Bartonella henselae</i>	Kyme <i>et al.</i> (2003)
<i>Bordetella bronchiseptica</i>	Irie <i>et al.</i> (2004), Irie <i>et al.</i> (2005), Irie <i>et al.</i> (2006), Mishra <i>et al.</i> (2005), Parise <i>et al.</i> (2007), Sloan <i>et al.</i> (2007)
<i>Bordetella parapertussis</i>	Uzureau <i>et al.</i> (2007)
<i>Brucella melitensis</i>	Boddey <i>et al.</i> (2006), Korbsrisate <i>et al.</i> (2005), Lee <i>et al.</i> (2010), Sawasdidoln <i>et al.</i> (2010), Taweechaisupapong <i>et al.</i> (2005), Tunpiboonsak <i>et al.</i> (2010)
<i>Burkholderia pseudomallei</i>	Fields and Thompson (2008), Gunther and Chen (2009), Hanning <i>et al.</i> (2008), Hanning <i>et al.</i> (2009), McLennan <i>et al.</i> (2008), Moe <i>et al.</i> , (2010), Murphy <i>et al.</i> (2006), Naito <i>et al.</i> (2010), Peyrat <i>et al.</i> (2008), Reeser <i>et al.</i> (2007), Reuter <i>et al.</i> (2010), Sulaeman <i>et al.</i> (2010), Svensson <i>et al.</i> (2009), Trachoo and Frank (2002), Trachoo <i>et al.</i> (2002)
<i>Campylobacter coli</i>	Varga <i>et al.</i> (2008)
<i>Campylobacter jejuni</i>	Olson <i>et al.</i> (2002)
<i>Clostridium perfringens</i>	Ballerig <i>et al.</i> (2009), Ciftci <i>et al.</i> (2009), Guiton <i>et al.</i> (2009), Macovei <i>et al.</i> (2009), Mohamed and Huang (2007), Oliveira <i>et al.</i> (2010), Teng <i>et al.</i> (2009)
<i>Corynebacterium pseudotuberculosis</i>	Shimoji <i>et al.</i> (2003)
<i>Corynebacterium renale</i>	Agladze <i>et al.</i> (2005), Beloin <i>et al.</i> (2008), Hancock <i>et al.</i> (2010), Olson <i>et al.</i> (2002), Prigent-Combaret <i>et al.</i> (2000), Puttamreddy <i>et al.</i> (2010), Uhlich <i>et al.</i> (2010), Wood (2009), Zogaj <i>et al.</i> (2001)
<i>Enterococcus faecalis</i>	Amer <i>et al.</i> (2010), Durham-Colleran <i>et al.</i> (2010), Margolis <i>et al.</i> (2010)
<i>Enterococcus faecium</i>	Jin <i>et al.</i> (2006, 2008)
<i>Erysipelothrix rhusiopathiae</i>	Olson <i>et al.</i> (2002), Sandal <i>et al.</i> (2007, 2009)
<i>Escherichia coli</i>	Ristow <i>et al.</i> (2008)
<i>Francisella novicida</i>	Amalaradjou <i>et al.</i> (2009), Gandhi and Chikindas (2007), Habimana <i>et al.</i> (2009), Harmsen <i>et al.</i> (2010a, 2010b), Latorre <i>et al.</i> (2010), Riedel <i>et al.</i> (2009), Takahashi <i>et al.</i> (2010), Todhanakasem and Young (2008)
<i>Francisella tularensis</i>	Olson <i>et al.</i> (2002)
<i>Haemophilus parasuis</i>	Carter <i>et al.</i> (2004), Cook <i>et al.</i> (2010), Johansen <i>et al.</i> (2009), Ojha <i>et al.</i> (2008), Wu <i>et al.</i> (2009), Yamazaki <i>et al.</i> (2006a, 2006b)
<i>Histophilus somni</i>	Daubenspeck <i>et al.</i> (2009), Justice-Allen <i>et al.</i> (2010), McAuliffe <i>et al.</i> (2006, 2008), Simmons and Dybvig (2007, 2009)
<i>Leptospira</i>	Olson <i>et al.</i> (2002)
<i>Listeria monocytogenes</i>	Bazire <i>et al.</i> (2010), Davies and Marques (2009), Deligianni <i>et al.</i> (2010), Fuxman Bass <i>et al.</i> (2010), Harmsen <i>et al.</i> (2010b), Lenz <i>et al.</i> (2008), Ma <i>et al.</i> (2009), Olson <i>et al.</i> (2002), Pérez-Osorio <i>et al.</i> (2010), Ryder <i>et al.</i> (2007)
<i>Mannheimia haemolytica</i>	Hu <i>et al.</i> (2010)
<i>Mycobacterium</i>	Jain and Chen (2007), Kim and Wei (2009), Marin <i>et al.</i> (2009), Olson <i>et al.</i> (2002), Römling (2005), Van Parys <i>et al.</i> (2010), Wong <i>et al.</i> (2010)
<i>Mycoplasma</i>	Boles <i>et al.</i> (2010), Dhanawade <i>et al.</i> (2010), Fox <i>et al.</i> (2005), Futagawa-Saito <i>et al.</i> (2006), Melchior <i>et al.</i> (2006a, 2006b, 2009), Nemati <i>et al.</i> (2009), Oliveira <i>et al.</i> (2006, 2007), Olson <i>et al.</i> (2002), Pérez <i>et al.</i> (2009), Tormo <i>et al.</i> (2005), Vancraeynest <i>et al.</i> (2004)
<i>Pasteurella multocida</i>	Bonifait <i>et al.</i> (2008), Grenier <i>et al.</i> (2009), Konto-Ghiorgi <i>et al.</i> (2009), Moscoso <i>et al.</i> (2009), Olson <i>et al.</i> (2002), Rinaudo <i>et al.</i> (2010), Tanabe <i>et al.</i> (2010), Wei <i>et al.</i> (2009)
<i>Pseudomonas aeruginosa</i>	Coquet <i>et al.</i> (2002), Darby (2008), Hinnebusch and Erickson (2008), Kim <i>et al.</i> (2008), Sun <i>et al.</i> (2009), Truchado <i>et al.</i> (2009), Wortham <i>et al.</i> (2010)
<i>Riemerella anatipestifer</i>	
<i>Salmonella</i>	
<i>Staphylococcus</i>	
<i>Streptococcus</i>	
<i>Yersinia</i>	

Treating Water...!



D Venne

Fig.81.36: Effect of dilution of a vaccine and Oxidation Reduction Potential (ORP) on the survival of Gumboro virus *in vitro*.



PREMISE

% on-farm compliance

 **SUCKS**

...or is not as good as you think





Biosecurity compliance Poultry Farms

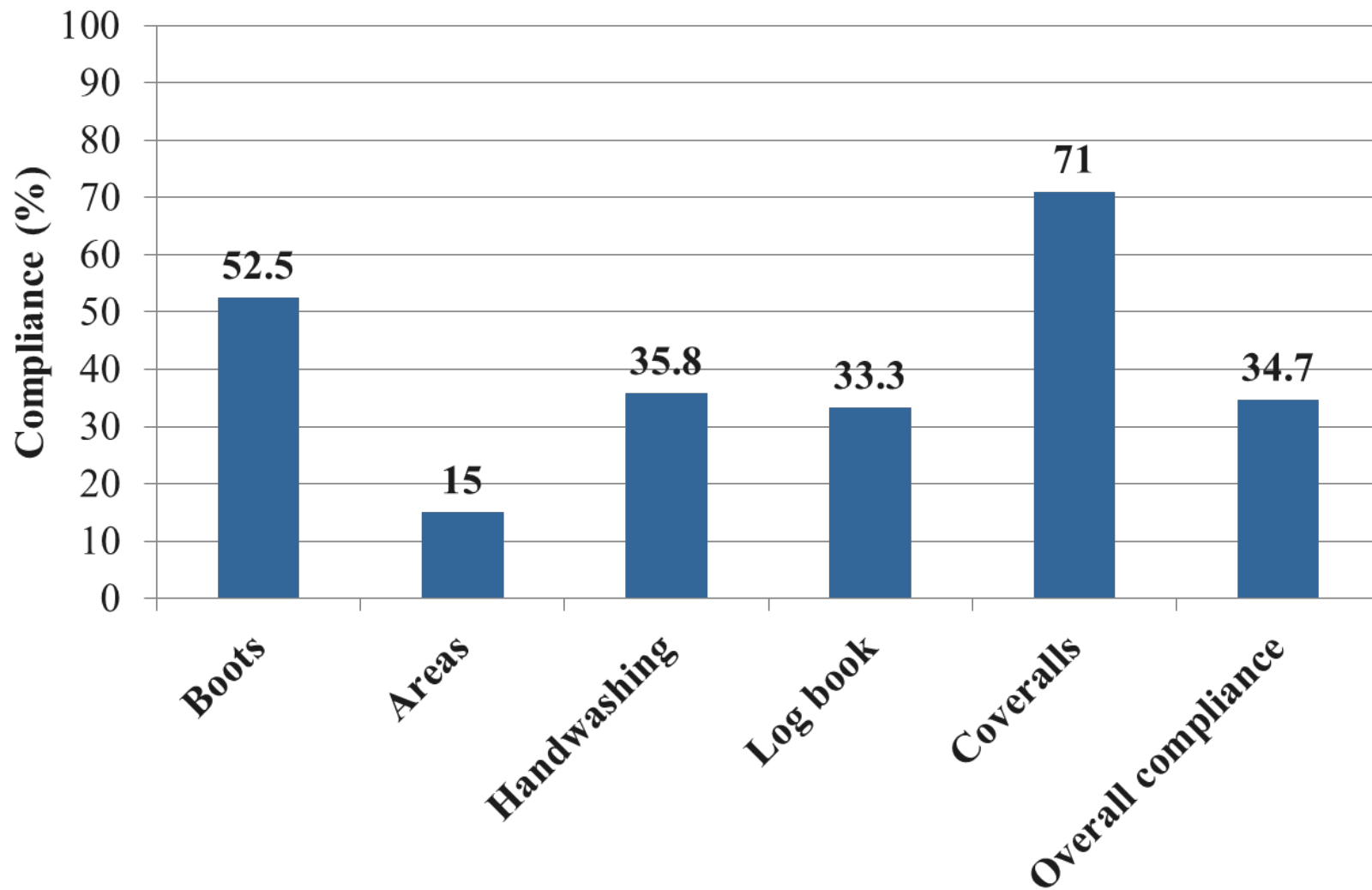
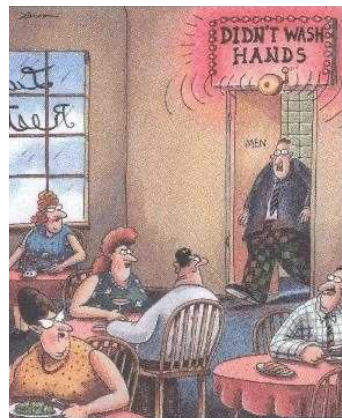
Description of 44 biosecurity errors while entering and exiting poultry barns based on video surveillance in Quebec, Canada

Manon Racicot^{a,b,*}, Daniel Venne^c, André Durivage^d, Jean-Pierre Vaillancourt^a



8 farms
883 visits
102 individuals

Human nature



Different situations require different measures

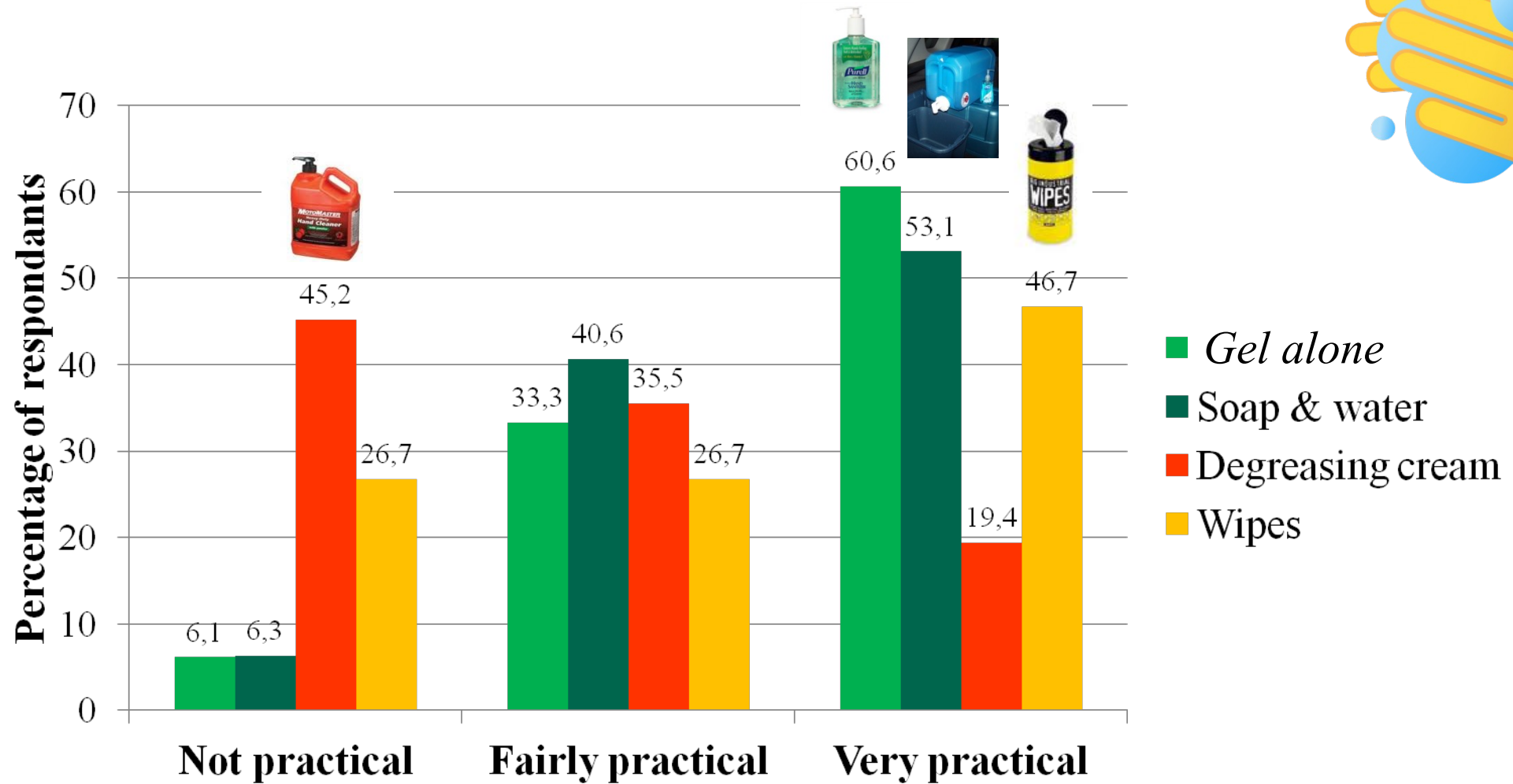


But there are universal elements to include in a biosecurity strategy

- Reach the right people
- Keep it simple
- Lead by example
- Communicate
- Provide feedback
- Incentive (reward)
- Make it easy: design
- Training - Simulations
- Buy-in: get all involved
- Innovate: technology

Human Perception & Beliefs

Practicality of protocols



Building design and farm layout

- Extremely large farms with multiple buildings
- Ponds/water bodies on farms
- Anything that requires regular and repeated entry to a building
- Open-sided buildings
- Uncontrolled fan outlets
- Inlets/fan with no biofilters
- Moving clean and dirty stuff in and out the same building entrance (cross-contamination)





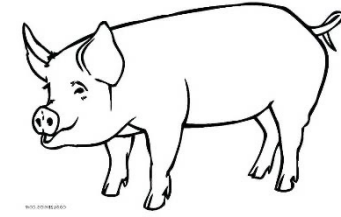
Mycoplasma: "Farm localization is the most important factor associated with reinfection....the second factor is the size of the neighboring farm"

RFW Goodwin, 1985

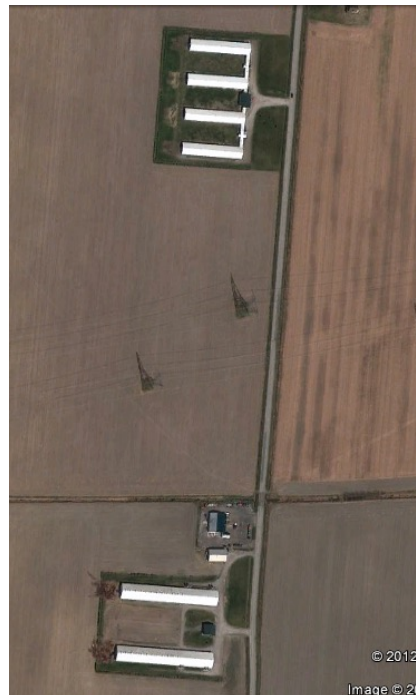




Regional farm density



Diseases	Risk factor	Risk level	Reference
Salmonella	High farm density	OR 2.2	Snow et al., 2012; Great Britain
Newcastle		OR 4.2	East et al., 2006; Australia
E. Coli		OR 6.3	Vandekerchove et al., 2004; Belgium
Avian influenza		OR 34.7	Boender et al., 2003, The Netherlands
PRRS		OR 7.3	Lambert et al., 2012, Canada



Less than 1 km (0.6 mile) between farms

- 2 x more chances → *Salmonella*
- 4 x more chances → Newcastle
- 6 x more chances → *E. Coli*
- 35 x more chances → A. influenza
- 7 x more chances → P.R.R.S.

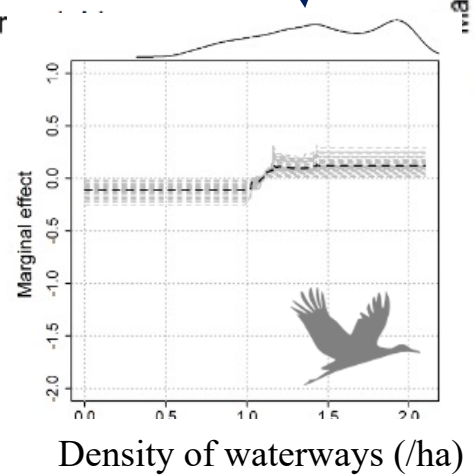
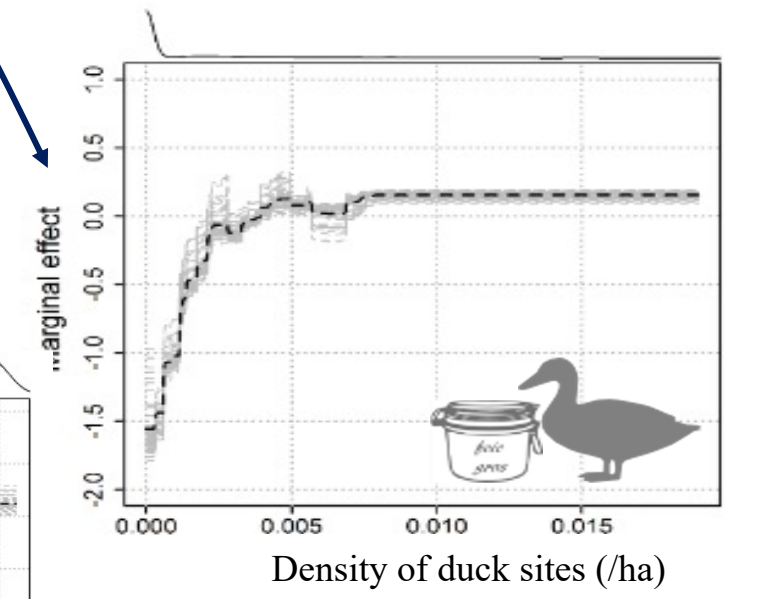
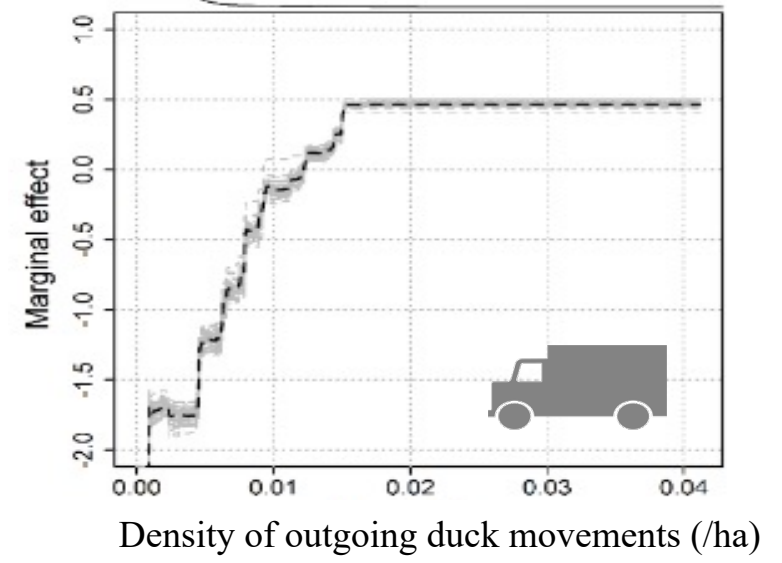
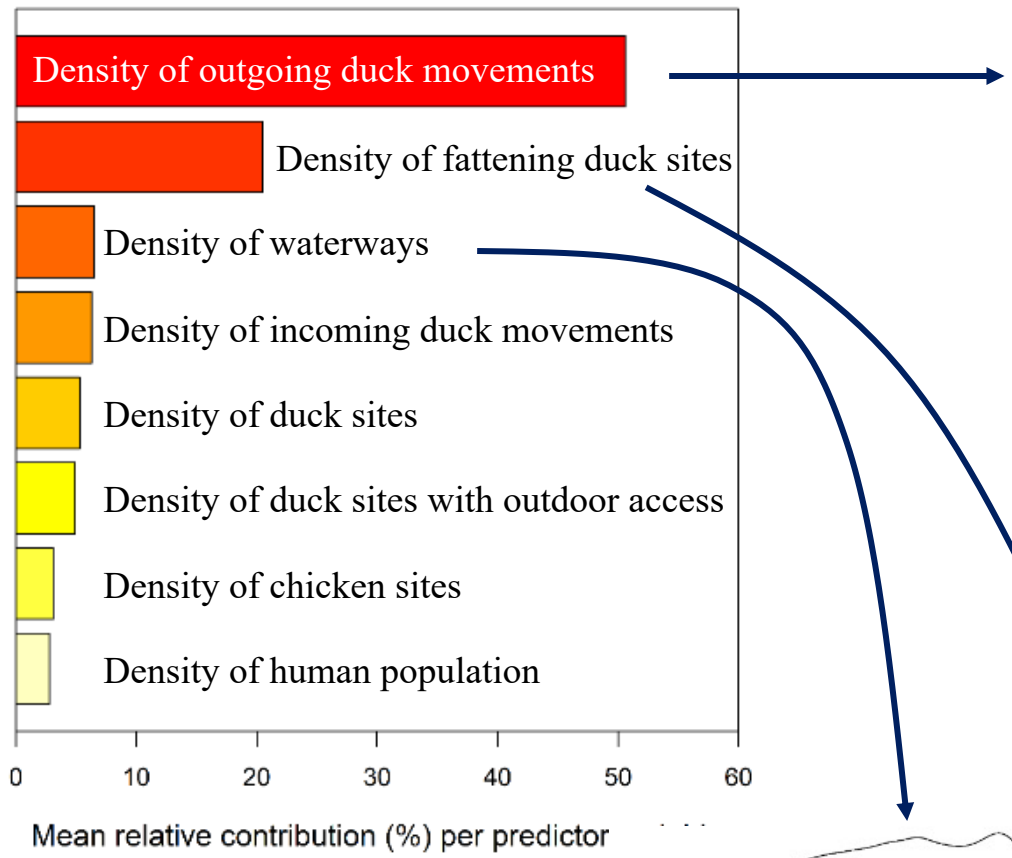
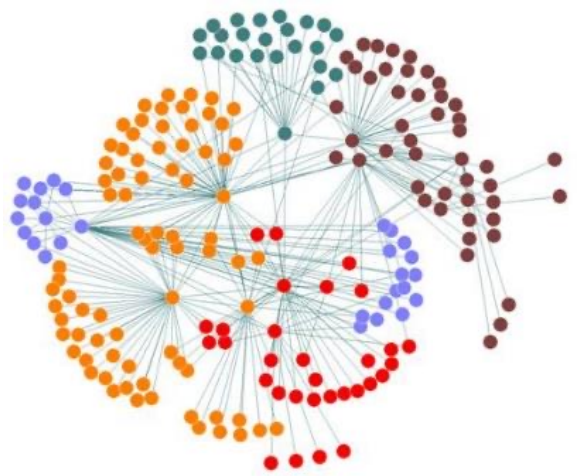
⇒ equipment, people, vehicles, wildlife, insects

Business → Farm 1 Farm 2 Farm 3

Accounting ≠ biology

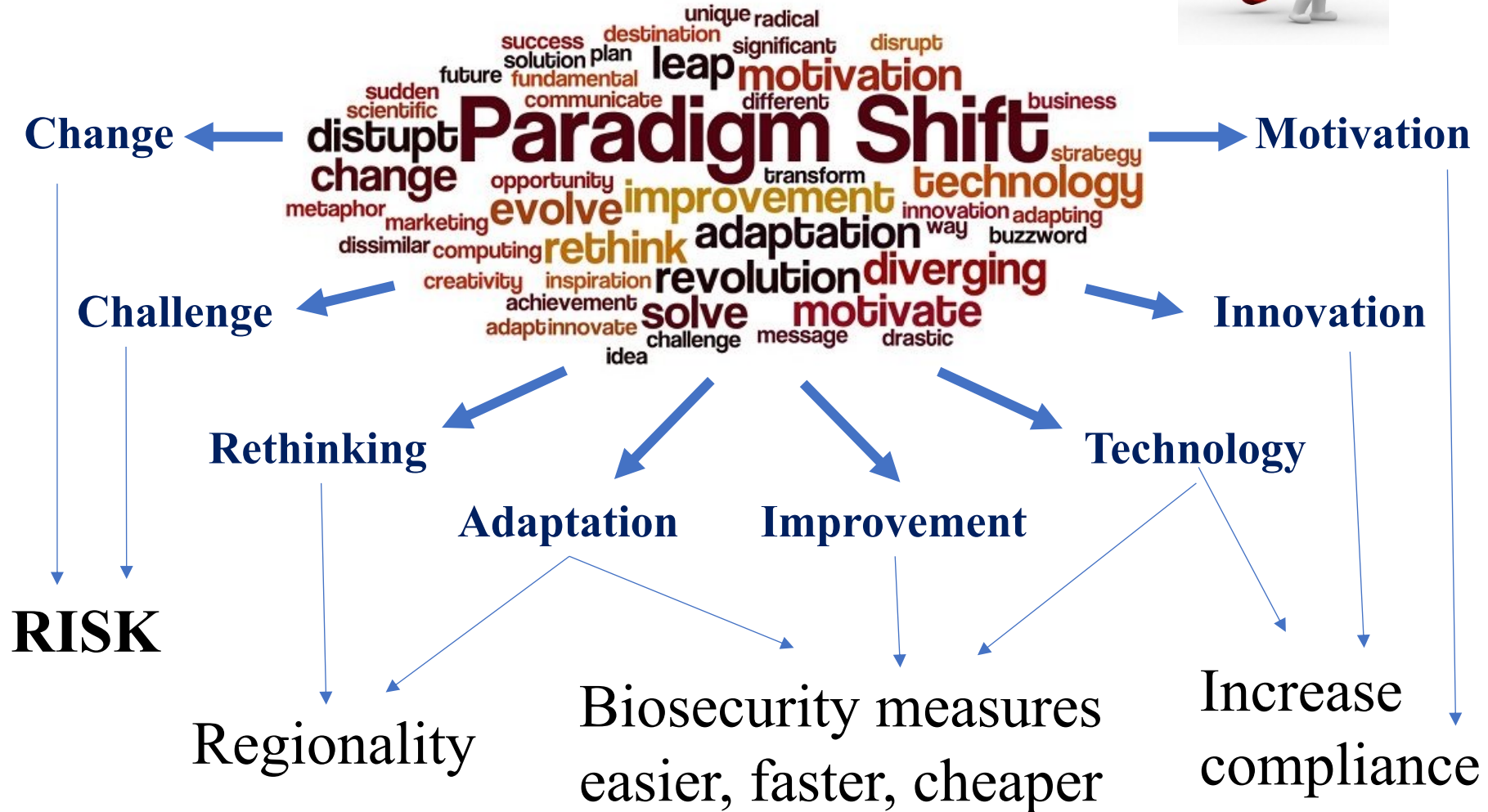
Nature (epidemiology) → One Big Fat Juicy Site!
(including viruses, bacteria, protozoa)

Hierarchy of explicative variables



Risk factors associated with highly pathogenic avian influenza in France – 2015-2017

What should we do?



What

Change

Challenge

RISK

Rethinking

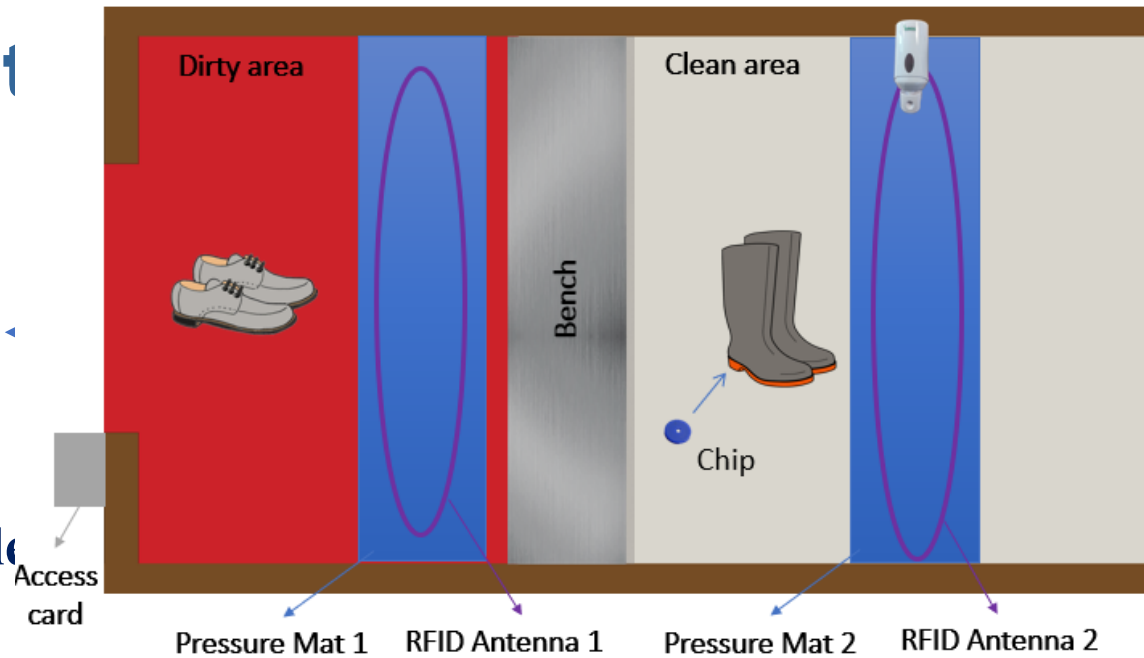
Regionality

Adaptation

**Biosecurity measures
easier, faster, cheaper**

Improvement

Technology



The pressure-sensitive strings play music while you go, and you can even download an “M-Pee-3” of your urinal masterpiece online afterwards.



What

Change

Chal

RISK

Need fixing/improvement:

- Movement of vehicles & personnel
- Zoning of production sites
- Rules for new constructions
- Dead bird management
- Anteroom (compliance)
- Cleaning & Disinfection
- Rodent control
- **Communication**

Adaptation

Improvement

Regionality

Biosecurity measures
easier, faster, cheaper

The pressure-sensitive strings play music while you go, and you can even download an “M-Pee-3” of your urinal masterpiece online afterwards.



Thank you!



Questions?